

**Adhiyamaan College of Engineering (Autonomous), Hosur**

**Department of Biotechnology**

**Academic year: 2019-20**

**1.1.3 Average percentage of courses having focus on employability/ entrepreneurship/ skill development offered by the institution**

<b>Program name</b>	<b>Course name</b>	<b>Course code</b>	<b>Activities/Content with direct bearing on Employability/ Entrepreneurship/ Skill development</b>	<b>Year of Introduction</b>
B.Tech Biotechnology	Pollution Control In Bioprocess Industries	515CHO05	Employability - The course offers skills like design and analysis of ideal and non -ideal reactors, also provides insight into the skills like Bioreactor assembly and manufacturing	2017-2018
B.Tech Biotechnology	Health & Pharmaceutical Biotechnology	615BTT04	Employability - The Course provides skills like design, manufacturing of drugs. The course also provides insight into the standard protocols to be followed in a drug manufacturing facility	2017-2018
B.Tech Biotechnology	Process Dynamics And Control	615EIO01	Employability - The course provides the students with the skill of basics of biological safety measures, industrial process dynamics and its control	2017-2018
B.Tech Biotechnology	Mini Project	715BTP08	Employability - The Project work provides the student with the skill set of managing project, planing and execution.	2018-2019
B.Tech Biotechnology	Employability Skills Lab	715BTP09	Employability - The course provides entrepreneurship-based skills like managing a firm, small business and to startups	2018-2019
B.Tech Biotechnology	Clinical Research And Database Management	715BTE01	Employability - The Course provides skills like design, manufacturing of drugs. The course also provides insight into the standard protocols to be followed in a drug manufacturing facility	2018-2019

B.Tech Biotechnology	Molecular Pathogenesis	815BTE03	Employability - Offers the student with therapeutics of diseases	2018-2019
B.Tech Biotechnology	Medical Coding	815BTE09	Employability - The course provides the students with the skill of basics of medical codes and transcripts	2018-2019
B.Tech Biotechnology	Project Work	815BTP05	Employability - The Project work provides the student with the skill set of managing project, planing and execution. In addition they provides the skill like report preparation, and presentation	2018-2019
B.Tech Biotechnology	Industrial Training	815BTP06	Employability - The Project work provides the student with the skill set of managing project, planing and execution. In addition they provides the skill like report preparation, and presentation	2018-2019

**ADHIYAMAAN COLLEGE OF ENGINEERING (AUTONOMOUS), HOSUR-635 109**

**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015  
Semester VI**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
615BTT04	HEALTH AND PHARMACEUTICAL BIOTECHNOLOGY	3	0	0	3	50	50	100

**Prerequisite** Biochemistry

*At the end of the course ,the students should be able to:*

- Course Objectives**
- *To have the basic knowledge of pharmacology*
  - *To gain knowledge in various dosage forms and bio pharmaceuticals*
  - *To be able to understand in pharmacokinetics and drug discovery*
  - *To understand the pharmaceutical dosage forms and applications*
  - *To gain the knowledge about the various biopharmaceuticals*

**UNIT I INTRODUCTION TO PHARMACOLOGY 9**

Historical outlines of drugs, classifications of drugs, physico-chemical properties of drugs, Routes of administration of drugs, drugs metabolism, controlled release drug delivery system, drug stability, Sources: plant, marine and microorganisms

**UNIT II DRUG DISCOVERY 9**

Drug discovery an introduction, basic clinical evolution of new drugs, bioavailability of drugs, quantitative and qualitative assay of drugs by biological testing, packing techniques like compression of tablets, wet & dry granulation, direct compression, tablet presses and coating

**UNIT III PHARMACOKINETICS AND BIOTRANSFORMATION 9**

Pharmacokinetics: introduction, absorption, distribution, elimination and metabolism of drugs, site of action, Phase I and Phase II reactions, prodrugs, adverse drug effects, Role of Enzymes in drug metabolism

**UNIT IV PHARMACEUTICAL DOSAGE FORMS AND APPLICATIONS 9**

Oral solid dosage forms, compressed tablets, types, pills, solutions, syrups, juices, nasal solutions, emulsions, lotions and extracts. Applications of various drugs in human body and site of action

**UNIT V BIO PHARMACEUTICALS 9**

Various categories of therapeutics like vitamins, laxatives, analgesics, contraceptives, common drugs which are abused, Antibiotics, human insulin, interferon, somatostalin, somatotropin - its preservation and analytical methods

**Total Hours 45**

*Upon Completion of this course, students will be able to get :*


<b>Course</b>	<i>CO1:Drugs, drugs action, drug metabolism</i>
<b>Outcomes</b>	<i>CO2:Various dosage forms of Biopharmaceuticals</i>
	<i>CO3:The recent evolution in pharmaceutical biotechnology</i>
	<i>CO4:evaluate different pharmaceutical parameters for the current and future biotechnology related products on the market.</i>
	<i>CO5: gained the knowledge about the various biopharmaceuticals</i>

**Text Books**

- 1 Remington, "The science and practice of pharmacy", Lippincott Williams and Wilkins, 20<sup>th</sup> edition, 2001
- 2 Gareth Thomas, Medicinal Chemistry an Introduction", John Wiley, New Delhi, 2000
- 3 Ram I. Mahato, Ajit S. Narang, "Pharmaceutical Dosage Forms and Drug Delivery", 2<sup>nd</sup> Edition, CRC Press, 2011
- 4 Mohsen A. Hedaya "Basic Pharmacokinetics", 2<sup>nd</sup> Edition, Routledge, 2012

**Reference Books**

- 1 Katzung, B.G. "Basic and Clinical Pharmacology", Prentice Hall of India, New Delhi., 1995
- 2 Tripathi, K.D. "Essentials of Medical Pharmacology", Jaypee Brothers Medical Publishers (P) Ltd, 6<sup>th</sup> edition, John Wiley, New Delhi, 2000



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**ADHIYAMAAN COLLEGE OF ENGINEERING (AUTONOMOUS), HOSUR-635 109**

**Department** BIOTECHNOLOGY      **Programme** B. TECH. - BT      **Regulation** 2015

**Semester VII**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		CA	EA	Total
715BTP09	EMPLOYABILITY SKILLS LAB	0	0	2	1	50	50	100

**Prerequisite** TECHNICAL ENGLISH I & II

**Course Objectives:**

- Course Objectives**
- To equip students of engineering and technology with effective speaking and listening skills in English.
  - To help them enrich their soft skills and interpersonal skills, which will make the transition from college to workplace smoother and help them excel in their career.
  - To enhance the performance of the students in the recruitment processes, self enhancement and launching start ups.

**Unit 1 Listening** 9

Listening Audios and Answering MCQs - Watching Video Clips on Famous Speeches, Motivational Videos, Documentaries and Answering MCQs - Listening Comprehension and TED talks.

**UNIT II Speaking** 9

Prepared Talk – Extempore - Story Knitting - Picture Talk – Brainstorming – Debates - Group Discussions - Elevator Speech - Mock HR Interviews - Story Narration – Miming - Short Skits.

**UNIT III Reading** 9

Reading Comprehension - Verbal Analogy - Classification - Alphabet Test - Logical Sequence of Words - Statement & Conclusions - Statement & Courses of Action - Situation Reaction Test - Theme Detection - Deriving Conclusions from Passages.

**UNIT IV Writing** 9

Business Letters - Email Writing (hints development) - Essay Writing - Paragraph Writing - Paraphrasing.

**UNIT V Career Skills** 9

Vocabulary Test (GRE, TOEFL, TOEIC & CAT Exam words) - Confused Pair of words - Contronyms - One Word Substitution - Sequencing of Sentences – Sentence Correction.

**Total Hours** 45

*On completion of the course, the students shall have the ability to:*

*CO1: Comprehend the various strategies of listening and its significance.*

*CO2: Articulate their views clearly and concisely with self-confidence and persuasiveness.*

**Course  
Outcomes**

*CO3: Understand the prevailing practices of testing in the recruitment process by the corporate and the institutional selection processes.*

*CO4: Communicate the corporate and social requirements in an impressive written mode.*

*CO5: Enhance their verbal skills in the screening tests competently both for recruitment and pursuing higher studies as well..*

**Text Books**

- 1 Agarwal R. S., A Modern Approach to Verbal and Non-verbal Reasoning, Chand & Co., New Delhi, 2012.

**Reference Books**

- 1 Lingua: Essays for TOEFL/IELTS, Dreamtech Press, New Delhi, 2016.
- 2 Lily Mangalam, Global English Comprehension, Allied Publishers Pvt. Ltd., New Delhi, 2014
- 3 Sharon Weiner Green and Ira K. Wolf, Barron's GRE, Glagotia Publications Pvt. Ltd., 18<sup>th</sup> Edition, New Delhi, 2011.
- 4 Mohamed Elias, R. Gupta's IELTS/TOEFL Essays, Ramesh Publishing House, 6<sup>th</sup> Edition, New Delhi, 2016.

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**Semester VII**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
715BTE01	CLINICAL RESEARCH AND DATABASE MANAGEMENT	3	0	0	3	50	50	100

**Prerequisite** Probability and statistics, Health and pharmaceutical technology

**Course Objective**

- Understand the drug development and study process through regulations
- To understand the roles and responsibilities of the clinical research team
- To know on review the CDM Start-up activities/documentation
- To know the view about the probability
- To gain knowledge related to research outcome

**UNIT I ETHICAL GUIDELINES 9**

Ethical Guidelines for Biomedical Research on Human guidelines – student of specific principles for clinical evaluation – Human Genome project - DNA banking – prenatal diagnosis – principles in transplantation. *regulatory affairs - GCP/ICH guidelines*

**UNIT II APPLICATIONS OF STATISTICS AND PROBABILITY 9**

Applications of Biostatistics in clinical Trial Management: Correlation - simple linear regression – multiple regressions – T-test - F-test – Chi square test - ANOVA – One way ANOVA. *Biostatistics and database management system.*

**UNIT III CONTRACT RESEARCHES 9**

Contract research – delivery model – CR Business environment – CR Information research – Contract research – Regulatory affairs of contract research – Clinical trial environment

**UNIT IV CLINICAL TRIALS OUT SOURCING 9**

Clinical trial – protocol approval – Informed consent – responsibility of sponsor – investigator – ethics committee – types of clinical trials – structure & contents of clinical report. Data blinding & Randomization – data management – trial subjects recruiting.

**UNIT V OUTSOURCING TRENDS-CASE STUDY OF MEDICAL CODING 9**

Introduction of medical coding and billing – Role of International classification of diseases book in medical coding- CPT (Current Procedure Terminology codes)-HIPAA (Health information portability and accounting act) - HCPCS (Healthcare Common Procedure Coding System)- CPC(Certified Professional Coder) –Medical billing and medical transcription-Medical coding job market in Business Process Outsourcing (BPO's) companies-starting own business sectors of medical coding and billing.

**Total Hours 45**

<b>Course</b>	<i>CO 1: Ability to describe about clinical research documentation and protocol</i>
<b>Outcomes</b>	<i>CO 2: knowledge on handling human and animal trials subjected to regulations</i> <i>CO 3: Knowledge on biostatistics subjected to validation on drug development</i> <i>CO4: knowledge related to clinical activities</i> <i>CO5: ability to describe about the database management</i>

#### **Text Books**

- 1 ICMR, "Ethical guidelines for biological research on human subjects", Indian Council of Medical Research Press, New Delhi, 2000.
- 2 International Classification of Diseases (ICD)- 10-CM, code Book diagnoses code set to assist in ICD- 10 training and code clarification, Tata Mc Graw Hill, New York, USA, 2012.
- 3 Knut Schoeder, "The 10 minutes Clinical Assessment", Wiley Black well, Singapore, 2010

#### **Reference Books**

- 1 The drug and cosmetic rule. Schedule Y., "Requirements and guidelines for permission to import and/ or manufacture of new drugs for sale or to undertake clinical trials". Government of India, New Delhi, 1945.
- 2 Machin, D. and Fayers, P., "Randomized clinical trials - Design, Practice and Reporting", Wiley Blackwell, Singapore, 2010.



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**Department BIOTECHNOLOGY Programme B. TECH. – BT Regulation 2015**  
**Semester VIII**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
815BTE03	MOLECULAR PATHOGENESIS	3	0	0	3	50	50	100

**Prerequisite** Basic Knowledge of Animal Biotechnology required

*At the end of the course ,the students should be able:*

- Course Objectives**
- To understand about the microbial toxins and modern molecular pathogenesis.
  - To know about the host pathogen interaction and identifying virulence factors.
  - To control pathogens by modern approaches.
  - To know about the pathogenic strategies
  - To understand the concept of the host defense mechanism

**UNIT I OVERVIEW**

9

Historical perspective - discovery of microscope, Louis Pasteur's contributions, Robert Koch's postulates, early discoveries of microbial toxins, toxic assays, vaccines, antibiotics and birth of molecular genetics and modern molecular pathogenesis studies, *Introduction to pathogenesis, components of microbial pathogenicity.*

**UNIT II HOST-DEFENSE AGAINST PATHOGENS AND PATHOGENIC STRATEGIES**

9

*Host defense against pathogens, clinical importance of understanding host defense, components of the host surface defenses systems like skin, mucosa, eye, mouth, respiratory tract, physical movements, limitation of free iron, antimicrobial compounds, mechanism of killing by humoral and cellular defense mechanisms, complements, inflammation process, general disease symptoms, Pathogenic adaptation to overcome the above defenses.*

**UNIT III MOLECULAR PATHOGENESIS (WITH SPECIFIC EXAMPLES)**

9

Virulence, virulence factors, virulence-associated factors and virulence lifestyle factors, molecular genetics and gene regulation in virulence of pathogens, molecular pathogenesis of Vibrio Cholerae: E.coli, Shigella, influenza virus, plasmodium.

**UNIT IV EXPERIMENTAL STUDIES ON HOST-PATHOGEN INTERACTIONS**

9

Virulence assays: adherence, invasion, cytopathic, cytotoxic effects. Criteria & tests in identifying virulence factors, attenuated mutants, molecular characterization of virulence factors, signal transduction & host responses, *virulence factors damaging the host tissues, virulence genes and regulation of the virulence genes.*

**UNIT V MODERN APPROACHES TO CONTROL PATHOGENS**

9

Classical approaches based on serotyping. Modern diagnosis based on highly conserved virulence factors, immuno & DNA-based techniques. New therapeutic strategies based on recent findings on

molecular pathogenesis of a variety of pathogens, Vaccines - modulation of immune response by vaccines, properties of vaccines, other immuno modulators.

**Total Hours 45**

**Course Outcomes**

*Upon Completion of this course, students will be able to get :*

*CO1: Knowledge Of Different Disease- Host Interactions Mechanisms In Organisms*

*CO 2: Concept About Modern Approaches To Control Pathogens*

*CO 3: Knowledge About Different Molecular-Molecular Pathogen Interaction*

*CO 4: Concept of different drug and pathogen interaction*


*CO 5: Idea of different vaccines to different pathogen*

**Text Books**

- 1 Clark V L and Bavoil P M, "Bacterial Pathogenesis", Academic Press, 1997.
- 2 Williams and Peter et al., "Bacterial Pathogenesis", (Methods in Microbiology Vol. 27), 1998.
- 3 Groisman and Eduardo A, "Principles of Bacterial Pathogenesis", Academic Press, 2001.
- 4 Nester, Anderson, Roberts, Pearsall, Nester, "Microbiology: A Human Perspective", 3<sup>rd</sup> Edition, McGraw-Hill, 2001.

**References**

- 1 Salyers, Abigail A and Dixie D. Whitt, "Bacterial Pathogenesis: A Molecular Approach", 2<sup>nd</sup> Edition, ASM, 2002.
- 2 McClane, Bruce A and Timothy A. Mietzner, "Microbial Pathogenesis: A Principles-Oriented Approach", Fence Creek Publishing, 1999.
- 3 Subramanian MA, "Toxicology: Principles and Methods", MJP Publishers, 2017.
- 4 "Bergey's Manual of Systematic Bacteriology", Vol. 1-3, 2nd Edition, Springer, 2005.



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Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
815BTE09	MEDICAL CODING	3	0	0	3	50	50	100
Prerequisite	NIL							

- Course Objectives**
- At the end of the course ,the students should be able to:*
- Develop comprehensive knowledge in the area of Human Anatomy & Physiology, Medical Coding, and CPT Coding.
  - Understand the knowledge of HCPCS Coding RCM, Coding Compliance, and HIPAA Laws.
  - Gain knowledge in anatomy
  - To improve skills in coding
  - Enhance the work activity related to physiology

**UNIT I HUMAN ANATOMY & PHYSIOLOGY PART I** 9

Cardiovascular System, Blood & its Components, Integumentary System, Endocrine System, Urology, Male Reproductive System. Location, Shape, Size, Structure, Physiology, Pathology, Diagnostic Test, Terminologies

**UNIT II HUMAN ANATOMY & PHYSIOLOGY PART II** 9

Female Reproductive Systems, Nervous System, Gastro Intestinal System, Pulmonology, Special Sciences, Orthopedics, Lymphatic System - Location, Shape, Size, Structure, Physiology, Pathology, Diagnostic Test, Terminologies

**UNIT III CURRENT PROCEDURE TERMINOLOGY CODING (CPT)** 9

CPT Codes, CPT Description, Medical Record Format, Speciality Listings and its Format, Usage of CPT Manuals, Software usage, Examples of CPT Speciality Code Practice, HCPCS Coding, Basic steps of HCPCS Coding, Differentiation of CPT and HCPCS Coding.

**UNIT IV INTERNATIONAL CLASSIFICATION OF DISEASE CODING (ICD)** 9

ICD Codes, ICD 9 CM – ICD 10 Transition, Diagnosis Interpretation, Usage of ICD Manuals, Index Listings, Tabular Listings, Software usage, Examples of Dx Code Practice.

**UNIT V MODIFIERS, E&M CODING, MEDICAL BILLING CYCLE & OVERVIEW** 9

Modifiers Listing, Usage and Indexing, E & M codes, classification, Application of E&M, Tabulation, Listings, Software usage, Examples of E&M Code Practice

**Total Hours 45**


- Course Outcomes**
- Upon Completion of this course, students will be able to get*
- Co1: Familiarize in the medical coding procedures for various treatment process. Co2: Acquire knowledge about ICD coding and medical billing process.
- Co3: Acquire knowledge about human anatomy & physiology. Co4: Familiarize in the software usage.
- Co5: Acquire knowledge about E&M Code Practice.

### **Text Books**

- 1 Current Procedural Terminology (CPT®) 2013 American Medical Association: I & II, Professional Edition (American Medical Association), CPT AMA Professional Edition, London, UK, 2013. ICD-9 CM Physicians Volume I and Volume II Contexo, A division of Access
- 2 Intelligence, London, UK, Medicine & Health Science Books, CPT 2009 Professional Edition, 2013.

### **References**

- 1 David N. Shier, Jackie Butler and Ricki Lewis, "Hole's Human Anatomy and Physiology Paperback – Import", McGraw Hill Higher Education, 12<sup>th</sup> edition, 2009.
- 2 Mader, "Understand Human Anatomy and Physiology Paperback", McGraw-Hill Education, 9<sup>th</sup> edition, 2006.
- 3 Carol J. Buck, "Step-by-Step Medical Coding 2014 Text + Workbook Paperback – Import", W B Saunders Co, CSM edition December, 2013.

  
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**Academic year: 2019-20**

**1.1.3 Average percentage of courses having focus on employability/ entrepreneurship/ skill development offered by the institution**

<b>Program name</b>	<b>Course name</b>	<b>Course code</b>	<b>Activities/Content with direct bearing on Employability/ Entrepreneurship/ Skill development</b>	<b>Year of Introduction</b>
B.Tech Biotechnology	Analytical Techniques In Biotechnology Lab	715BTP10	Entrepreneurship - The course will help the students to enhance their language skills to the next level and will hone their presentation skills, active participation in GD, critical thinking and leadership skills.	2018-2019
B.Tech Biotechnology	Bioethics, IPR And Entrepreneurship	815BTT01	Entrepreneurship - The course provides entrepreneurship-based skills like managing a firm, small business and to startups. The course provides the student in promoting the Entrepreneurship with the basics of IPR generation and filing, biobusiness management and group presentation among the peers	2018-2019

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Semester VI**

Course Code	Course Name	Hours/Week				Credit	Maximum Marks		
		L	T	P	C		CA	EA	Total
715BTP10	<b>ANALYTICAL TECHNIQUES IN BIOTECHNOLOGY LAB</b>	0	0	1	0		50	50	100

**Prerequisite** Instrumental Methods of Analysis Lab

*At the end of the course, the students should be able to:*

**Course** • Develop skills and techniques used in modern biotechnology.

**Objectives**

- Learn the techniques of chromatography
- Gain knowledge related to the hybridization techniques
- Ability to develop the techniques of fermentor types
- Develop skills about the types of centrifugation

**LIST OF EXPERIMENTS**

1. Various types of Centrifugation
2. Running of a pilot fermentor
3. 2D gel Electrophoresis
4. ELISA
5. DNA Hybridization
6. Isoelectric Focusing
7. Electroporation
8. High Performance Liquid Chromatography
9. Thin Layer Chromatography
10. Vermicomposting
11. COD Analyser

**Total hours 45**

*Upon Completion of this course, students will be able to get :*

**Course Outcomes** CO1: Knowledge on practical skills in analytical techniques and instrumentation of biotechnology

CO2: Analytical skills to do project

CO3: Learn various separation techniques involved in biotechnology industries

CO4: Student develop the techniques knowledge of fermentor types

CO5: Developed skills about the types of centrifugation


**Text Books**

Keith Wilson and John Walker, Practical Biochemistry – Principles and techniques, Cambridge

1 University Press, U.K; 5th Edition, 2003

2 Frank C. Hay, Olwyn M.R. Westwood, "Practical

Immunology; Blackwell Science; 4th edition (January 28, 2002)

  
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**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015**

**Semester VIII**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
815BTT01	BIOETHICS, IPR AND ENTREPRENEURSHIP	3	0	0	3	50	50	100

**Prerequisite**

*At the end of the course, the students should be able to:*

- Course Objectives**
- To create awareness about IPR and Engineering ethics
  - To follow professional ethics and practices in their careers
  - To create awareness and responsibilities about the environment and society
  - To learn the present work related to trade marketing
  - To create the view with respect to ethics in biotechnology

**UNIT I HISTORY OF BIOETHICS**

9

Bioethics as a discipline – philosophical reflections on experimenting with human subjects - active and passive euthanasia; culture assumption in the history of Bioethics– medical ethics in India and America.

**UNIT II METHODS OF ETHICAL ANALYSIS**

9

Ethical reasoning- philosophical, clinical and cultural dimensions; challenge of ethical relativism; methods of philosophical theories and principles- Equality and its implications; methods of casuistry and methods of narrative approaches

**UNIT III ETHICS IN BIOTECHNOLOGY**

9

Ethics committee (hospital) – Inner working of an ethics committee; ethics consultation – skills, roles and training; Biosafety regulation-national and International guidelines; rDNA guidelines-guidelines for rDNA research activities, mechanism of implementation of biosafety guidelines

**UNIT IV PATENTING, IPR AND APPLICATIONS**

9

Introduction to Intellectual property rights, types: patents, copy right, trade mark, trade secret, geographical indications, importance of IPR, Patenting and non patenting life, TRIPS

**UNIT V ENTREPRENEURSHIP IN BIOTECHNOLOGY**

9

The Significance of the Biotechnology Entrepreneur; The Integration of Two Distinctly Different Disciplines Biotechnology Entrepreneurship Versus General Entrepreneurship ; Entrepreneurship and Intrapreneurship Essential Biotechnology Entrepreneurial Characteristics; Four Backgrounds of Biotechnology Entrepreneurs

**Total Hours 45**

*Upon Completion of this course, students will be able to :*

**Course  
Outcomes**

*CO: 1 Touches on fundamental values, such as human dignity and the genetic integrity of humanity.*

*CO:2 Serve basic human needs such as human health, food and a safe environment, CO:3 Raise human rights issues such as access to health and benefits from scientific progress*

*CO: 4 Concerns over equitable access to the fruits of new technologies, the consent of those involved in research, and protection of the environment.*

*CO:5 Obtaining a clear information on the entrepreneurship and understand their economic values*

**Text Books**


- 1 Bioethics , second edition , Nancy S.Jecker , Albert R.Jonsen,RobertA,Pearlman.Jones and Bartlett Publishers,2003.
- 2 Singh K, “ Intellectual Property Rights on Biotechnology”, BCIL, New Delhi,2001.
- 3 M.K. Sateesh, “Bioethics and Biosafety”, I.K. International Publishing House pvt. Ltd, 2008.

**References**

- 1 Entrepreneurship Development – Poornima.M.Charantimath – Small Business Enterprises – Pearson Education – 2006
- 2 Sasson A, “ Biotechnologies and Development”, UNESCO Publications, 1998
- 3 Sasson A, “ Biotechnologies in Developing countries present and future”, UNESCO Publishers, 1993

**E-Books**

- 1 Biotechnology Entrepreneurship: Starting, Managing, and Leading Biotech by Craig Shimasaki

  
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B.Tech Biotechnology	Technical English	118ENT01	Skill Development - This course enhances increase students' efficiency in their academic and general reading, pronunciation in real-lifesituations, pronunciation in real-lifesituations and augmenting the technical writing skills like writing letters in formal and business situations	2018-2019
B.Tech Biotechnology	Engineering Mathematics-I	118MAT02	Skill Development - This course enable basic skills on the eigen value problems and differential equations of certain types, including systems of differential equations	2018-2019
B.Tech Biotechnology	Engineeirng Physics	118PHT03	Skill Development - This course provides skills on the concept of properties of matter, the properties of sound and principles of quantization of energy and coherent light and its importance	2018-2019
B.Tech Biotechnology	Engineering Chemistry	118CYT04	Skill Development - The students can understand and apply the concepts in electrochemistry and Energy storage devices, the chemistry of Corrosion, concepts of thermodynamics and phase equilibrium	2018-2019

B.Tech Biotechnology	Engineering Graphics	118EGT05	Skill Development - The students will learn graphics skill for communication of concepts, ideas and design of engineering products and expose them to existing national standards related to technical drawings.	2018-2019
B.Tech Biotechnology	Engineering Chemistry Laboratory	118CYP07	Skill Development - The students can understand and apply the concepts in electrochemistry and Energy storage devices, the chemistry of Corrosion, concepts of thermodynamics and phase equilibrium	2018-2019
B.Tech Biotechnology	Engineering Practice Laboratory	118EPP08	Skill Development - This course enable the student to learn about the basics of computer and problem solving methods.	2018-2019
B.Tech Biotechnology	Basic Civil and Mechanical Engineering	118ESE01	Skill Development - This course enable students to gain the knowledge on civil works like masonry, roofing, flooring and plastering	2018-2019
B.Tech Biotechnology	Communicative English	218ENT01	Skill Development - This course facilitate students amplify suitable language skills for academic and professional purposes, vocabulary power, different functions of technical and scientific English	2018-2019
B.Tech Biotechnology	Engineering Mathematics-II	218MAT02	Skill Development - This course facilitates double and triple integration concepts and apply to study vector calculus comprising of surface and volume integrals along with the classical theorems involving them.	2018-2019
B.Tech Biotechnology	Environmental Science and Engineering	218GET03	Skill Development - This course enable students study the nature and facts about environment	2018-2019
B.Tech Biotechnology	Engineering Mechanics	218EMT04	Skill Development - This course make the students understand the vectorial and scalar representation of forces and moments, static equilibrium of particles and rigid bodies both in twodimensions	2018-2019
B.Tech Biotechnology	Problem Solving and Python Programming	218PPT05	Skill Development - This course enable the student to learn about the basics of computer and problem solving methods.	2018-2019

B.Tech Biotechnology	Biochemistry	318BTT02	Skill Development - This course provides students to get skill in various metabolic pathways and its regulation	2019-2020
B.Tech Biotechnology	Cell Biology	318BTT03	Skill Development - This course enables students skill in cellular signalling mechanisms, cellular regulations and cell culture techniques	2019-2020
B.Tech Biotechnology	Microbiology	318BTT04	Skill Development- This course provides skills in microbial classification, identification and control	2019-2020
B.Tech Biotechnology	Instrumental Methods of Analysis	318BTT05	Skill Development - This course provides basic skills on biological instrumentation	2019-2020
B.Tech Biotechnology	Basic Industrial Biotechnology	318BTT06	Skill Development - This course provides skills on production of biologically important products such as antibiotics, vitamins, alcohol, etc.,	2019-2020
B.Tech Biotechnology	Biochemistry Lab	318BTP07	Skill Development - This course provides basic skills on qualitative and quantitative identification of biomolecules	2019-2020
B.Tech Biotechnology	Cell Biology Lab	318BTP08	Skill Development - This course provides basic skills on identification of cellular mechanisms	2019-2020
B.Tech Biotechnology	Microbiology Lab	318BTP09	Skill Development - This course enables skills on microbial culture techniques and identification of microorganisms	2019-2020
B.Tech Biotechnology	Probability And Statistics	418PBT01	Skill Development - This course enables skills on design of experiments and research methodologies	2019-2020
B.Tech Biotechnology	Molecular Biology	418BTT02	Skill Development - This course provide skills on genomic and plasmid DNA and RNA isolation, PCR based techniques and cloning and expression of vectors	2019-2020
B.Tech Biotechnology	Stoichiometry And Process Calculations	418BTT03	Skill Development - This course provide skills on stoichiometric calculations for various industrial operations	2019-2020
B.Tech Biotechnology	Fundamentals of Unit Operations	418BTT04	Skill Development - This course provide skills on bioreactor and heat transfer operations	2019-2020

B.Tech Biotechnology	Enzyme Technology	418BTT05	Skill Development - This course provide skills on industrially important enzyme production and activity determination	2019-2020
B.Tech Biotechnology	Environmental Biotechnology	418BTT06	Skill Development - This course provide skills on environmental studies, bioremediation and waste management	2019-2020
B.Tech Biotechnology	Molecular Biology Lab	418BTP07	Skill Development - This course provide skills on genomic and plasmid DNA and RNA isolation, PCR based techniques and cloning and expression of vectors	2019-2020
B.Tech Biotechnology	Instrumental Methods of Analysis Lab	418BTP08	Skill Development - This course provides basic skills on biological instrumentation	2019-2020
B.Tech Biotechnology	Enzyme Technology Lab	418BTP09	Skill Development - This course provide skills on industrially important enzyme production and activity determination	2016-2015
B.Tech Biotechnology	Immunology	515BTT01	Skill Development - The course provides the students with the skillset of raising antigen and antibodies against the various disease, and the detection procedure.	2017-2018
B.Tech Biotechnology	Genetic Engineering	515BTT02	Skill Development - This course provide skills on genomic and plasmid DNA and RNA isolation, PCR based techniques and cloning and expression of vectors	2017-2018
B.Tech Biotechnology	Bioprocess Engineering I	515BTT03	Skill Development - The course offers the students with the skill set of handling equipment related to heat transfer. In addition, the course provides the students with an deeper insight into the diffusion, distillation, adsorption, extraction and leaching which is most important skills as a chemical process engineer	2017-2018

B.Tech Biotechnology	Fundamentals of Mass Transfer	515BTT04	Skill Development - The course offers the students with the skill set of handling equipment related to heat transfer. In addition, the course provides the students with an deeper insight into the diffusion, distillation, adsorption, extraction and leaching which is most important skills as a chemical process engineer	2017-2018
B.Tech Biotechnology	Chemical Thermodynamics & Biothermodynamics	515BTT05	Skill Development - The course offers the students with the skill set of handling equipment related to heat transfer. The course offers a insight into the basic skill sets like understanding the thermodynamics of the reaction, thermodynamics related to the microbial growth and the product formation	2017-2018
B.Tech Biotechnology	Immunology Lab	515BTP07	Skill Development - The course provides the students with the skillset of raising antigen and antibodies against the various disease, and the detection procedure.	2017-2018
B.Tech Biotechnology	Genetic Engineering Lab	515BTP08	Skill Development - The course provides the students with the skill set of generating a recombinant DNA, cloning and expression of vectors, genome mapping and sequencing and PCR based skills	2017-2018
B.Tech Biotechnology	Bioprocess Engineering Lab I	515BTP09	Skill Development - The course offers a clear insight into the basic skills required for a Bioprocess Engineers. The course offers the students with the skill set of handling equipment related to chemical process engineer	2017-2018
B.Tech Biotechnology	Bioinformatics	615BTT01	Skill Development - The course provides the students with the skill of basics of database in biological system, sequence alignment, phylgogeny and CADD	2017-2018
B.Tech Biotechnology	Chemical Reaction Engineering	615BTT02	Skill Development - The course offers skills like design and analysis of idelal and non -ideal reactors, also provides insight into the skills like Bioreactor assembly and manufacturing	2017-2018

B.Tech Biotechnology	Bioprocess Engineering-II	615BTT03	Skill Development - The course focusses on the preparation, sterilization of media, design and optimization of media. Understand the basic of the different kinds of the fermenter and the kinetics of product formation	2017-2018
B.Tech Biotechnology	Plant Biotechnology	615BTT05	Skill Development - The course is focussed in the manner to provide indepth application oreinted skills like gene transfer in plants, transgenic and commercialization	2017-2018
B.Tech Biotechnology	Bioprocess Engineering Laboratory -II	615BTP07	Skill Development - The course focusses on the preparation, sterilization of media, design and optimization of media. Understand the basic of the different kinds of the fermenter and the kinetics of product formation	2017-2018
B.Tech Biotechnology	Plant Biotechnology Laboratory	615BTP08	Skill Development - The course is focussed in the manner to provide indepth application oreinted skills like gene transfer in plants, transgenic and commercialization	2017-2018
B.Tech Biotechnology	Technical Seminar	615BTP09	Skill Development - The course will help the students to enhance their language skills to the next level and will hone their presentation skills, active participation in GD, critical thinking and leadership skills.	2017-2018
B.Tech Biotechnology	Animal Biotechnology	715BTT01	Skill Development - The course provides the students with the skill of animal diseaes, treatment, micromanipulation and transgenics animal technology	2018-2019
B.Tech Biotechnology	Downstream Processing	715BTT02	Skill Development - The course provides the students with the skill of choice of different downstream processing like filtration, chromatography, and othe size-based methods	2018-2019
B.Tech Biotechnology	Cancer Biology	715BTT03	Skill development - This course enables students skill in cellular signalling mechanisms, cancer cell proliferation and regulations and control mechanisms	2018-2019

B.Tech Biotechnology	Disaster Management	715BTT04	Skill Development - Aseismic design is mandatory as per IS recommendations. This course develops the skill set required for aseismic design of structures	2018-2019
B.Tech Biotechnology	Downstream Processing Lab	715BTP07	Skill Development - The course provides the students with the skill of choice of different downstream processing like filtration, chromatography, and other size-based methods	2018-2019
B.Tech Biotechnology	Genomics And Proteomics	715BTE10	Skill Development - The course provides the students with the skill of genome and proteome analysis	2018-2019

**OBJECTIVES**

At the end of the course, the students should be able to:

- To develop strategies and skills to enhance their ability to read and comprehend engineering and technology texts.
- To foster the ability to write convincing job applications and effective reports.
- To develop their speaking skills to make technical presentations, participate in group discussions.
- To strengthen their listening skills which will help them comprehend lectures and talk in their area of specialization.

**UNIT I**

9

**Listening** - Ink talks and gap exercises - **Speaking** – Asking for and giving directions - **Reading** – short technical texts from journals and newspapers - **Writing** - definitions – instructions – checklists – recommendations - **Vocabulary Development** - technical vocabulary - **Language Development** – parts of speech – articles – word formation.

**UNIT II**

9

**Listening** - longer technical talks - **Speaking** – process description - **Reading** – longer technical texts – **Writing** – graphical representation - **Vocabulary Development** - vocabulary used in formal letters/emails and reports – **Language Development** – tenses - voices - numerical adjectives – question tags.

**UNIT III**

9

**Listening** - listening to classroom lectures - **Speaking** – introduction to technical presentations - **Reading** – longer texts both general and technical and practice in speed reading – **Writing** – process description using sequence words and sentences - **Vocabulary Development** - Misspelled words – one-word substitution - **Language Development** - embedded sentences – singular and plural nouns - compound nouns - editing

**UNIT IV**

9

**Listening** - Listening to documentaries and making notes - **Speaking** – mechanics of presentations - **Reading**– reading comprehension – **Writing** - email etiquettes - job application – cover letter –Résumé preparation - essay writing - **Vocabulary Development** – synonyms and antonyms –paraphrasing - **Language Development** – modals– conditionals.

**UNIT V**

9

**Listening** - TED talks - **Speaking** – brainstorming and debate – **Reading** – reading and understanding technical articles – **Writing** – reports - minutes of a meeting - **Vocabulary Development**- verbal analogies - phrasal verbs - **Language Development** - concord - reported speech.

**Total Hours 45 PERIODS**

**COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

CO1: Read technical texts and write area- specific texts effortlessly.



CO2: Listen and comprehend lectures and talks in their area of specialization successfully.

CO3: Speak appropriately and effectively in varied formal and informal contexts.

CO4: Understand the basic grammatical structures and its applications.

#### **TEXT BOOKS**


1. Board of editors. Fluency in English A Course book for Engineering and Technology. OrientBlackswan, Hyderabad: 2016.
2. Sudharshana. N. P and Saveetha. C. English for Technical Communication. (
3. Uttham Kumar. N. Technical English I (with work book). Sahana Publications, Coimbatore, 2016.

#### **REFERENCE BOOKS**

1. Raman, Meenakshi and Sharma, Sangeetha- Technical Communication PrinciplesandPractice. Oxford University Press: New Delhi,2014.
2. Kumar, Suresh. E. Engineering English. Orient Blackswan: Hyderabad,2015.
3. Booth-L. Diana, Project Work, Oxford University Press, Oxford: 2014.
4. Grussendorf, Marion, English for Presentations, Oxford University Press, Oxford: 2007
5. Means, L. Thomas and Elaine Langlois, English & Communication for Colleges. CengageLearning, USA: 2007.

**Note:** The book given under Extensive Reading is meant for inculcating the reading habit of the students.

They need not be used for testing purposes.



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**OBJECTIVES**

At the end of the course, the students should be able to:

- To understand the eigen value problems.
- To solve differential equations of certain types, including systems of differential equations that they might encounter in the same or higher semesters.
- To understand the concepts of curvatures, evolutes and envelopes and to study the maxima and minima of any function.
- To learn the partial derivations and apply the same to find maxima and minima.
- To solve certain linear differential equations using the Laplace transform technique which has applications in control theory and circuit theory.

**UNIT I** 9

Eigen values and eigen vectors of a real symmetric matrix – Properties – Cayley - Hamilton theorem (Statement only) – Orthogonal transformation of a symmetric matrix to diagonal form – Quadratic form – Reduction of quadratic form to canonical form by orthogonal transformation.

**UNIT II** 9

Curvature in Cartesian co-ordinates – Centre and radius of curvature – Circle of curvature – Evolutes – Envelopes – Evolutes as envelope of normals.

**UNIT III** 9

Partial derivatives – Euler's theorem for homogenous functions – Total derivatives – Jacobians – Taylor's expansion – Maxima and Minima – Method of Lagrangian multipliers.

**UNIT IV** 9

Higher order linear differential equations with constant coefficients – Method of variation of parameters

– Cauchy's and Legendre's linear equations – Simultaneous first order linear equations with constant coefficients – Applications to Engineering Problems – Electric Circuits, Simple Harmonic Motions and bending of beams.

**UNIT V** 9

Laplace transforms – Conditions for existence – Basic properties (without proof) – Laplace Transform of elementary functions, derivatives and integrals, unit step function and impulse functions, periodic functions. Definition of Inverse Laplace transform – Convolution theorem (Statement and applications only) – Initial and final value theorems (Statement and applications only) – Solution of linear ordinary differential equations of second order with constant coefficients using Laplace transform techniques.

**TOTAL HOURS 45 PERIODS**

**COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

CO1: Develop the knowledge of basic linear algebraic concepts.

CO2: Determine the solutions of ordinary differential equations by various methods which have an application in their core subjects.

CO3: Acquire the basic knowledge of ordinary differential calculus. CO4: Compute maxima and minima of a function.


CO5: Apply Laplace transform techniques to solve ordinary differential equations which have an application in many engineering fields

#### **TEXT BOOKS**

1. Erwin Kreyszig, "Advanced Engineering Mathematics", Wiley India, 10<sup>th</sup> edition New Delhi 2016.
2. Grewal. B.S, "Higher Engineering Mathematics", 43rd Edition, Khanna Publications, Delhi, 2014.

#### **REFERENCE BOOKS**

1. T.Veerarajan, "Engineering Mathematics " Tata McGraw-Hill Publishing company, New Delhi, 2014.
2. Kandasamy.P, Thilagavathy,K., & Gunavathi.K., "Engineering Mathematics for first year ", S.Chand &Company Ltd., New Delhi,2014.
3. Ramana B.V., "Higher Engineering Mathematics", Tata McGraw Hill Co. Ltd., New Delhi, 11<sup>th</sup> Reprint, 2010.

  
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**OBJECTIVES**

At the end of the course, the students should be able to:

- To understand the concept of properties of matter.
- To understand the properties of sound and principles of quantization of energy.
- To understand the properties of coherent light and its importance.

**UNIT I PROPERTIES OF MATTER 9**

Elasticity – Stress – Strain diagram – Factors affecting elasticity – Twisting couple on a wire – Torsion pendulum – Young’s modulus - cantilever - Heavy cantilever – Uniform and Non uniform bending (theory and experiment)–Viscosity- Poiseuille’s method for Coefficient of Viscosity.:

**UNIT II ACOUSTICS AND ULTRASONICS 9**

Classification of sound, loudness, intensity – Decibel – Weber Fechner Law – Reverberation and Reverberation time –derivation of Sabine’s formula for Reverberation time (Growth and Decay)–Absorption coefficient and itsdetermination. Introduction of Ultrasonics – Production – magnetostriction effect – magnetostriction generator – piezoelectric effect–piezo electric generator–Detection of ultrasonic waves, properties–Cavitation–Applications–Depth of sea – Non Destructive Testing.

**UNIT III QUANTUM PHYSICS 9**

Black body radiation–Planck’s theory (derivation)–Deduction of Wien’s displacement law and Rayleigh–jeans’ Law from Planck’s theory – Compton Effect – derivation – Matter waves – Schrödinger’s wave equation – Time independent and time dependent equations– Physical significance of wave function – Particle in a one dimensional box – Degeneracy and Non Degeneracy.

**UNIT IV LASER 9**

Introduction – Principle of Spontaneous emission and stimulated emission – Population inversion – pumping– Einstein’s A and B coefficients – derivation – Types of lasers – He-Ne, CO<sub>2</sub>, Nd-YAG, Semiconductorlasers – homojunction – Applications of Laser.

**UNIT V WAVE OPTICS & FIBRE OPTICS 9**

Interference – Air wedge (theory & experiment) – Polarization– Methods of polarizing light– Theory of plane circularly and elliptically polarized light. Principle and propagation of light in optical fibers – Numerical aperture and Acceptance angle – Types of optical fibers (material, refractive index, and mode) – Fiber optical communication system (Block diagram) – Fiber optic sensors – Temperature & Displacement sensors (Qualitative).

**TOTAL HOURS 45 PERIODS**

**COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

CO1: To understand properties of solids with different types of moduli and to gain knowledge about absorption coefficients of solids and different surfaces.

CO2: To understand basic concepts of high frequency sound waves and its applications.

CO3: To understand basic concepts of quantum mechanical behavior of wave and particle along

with applications.

CO4: To understand the concepts of production of laser and its behavior with diffraction principle of interference.

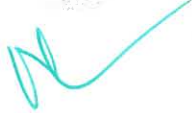
CO5: To apply the concept of polarization phenomenon and thereby its applications in fiber optic communication.

**TEXT BOOKS**

1. R.K. Gaur and S.C. Gupta, 'Engineering Physics' Dhanpat Rai Publications, New Delhi (2003)
2. Jayaprakash R.N, 'Engineering Physics - I', Dhanam Publications, Chennai, (2007).

**REFERENCE BOOKS**

1. R. Murugesan ,Kiruthiga Sivaprasath , Modern Physics S. Chand publications 2016,New Delhi.
2. GhatakOptics the McGraw-Hill Companies, Inc., 1221 Avenue of the Americas, New York, NY 10020.
3. Dr.M.N.Avadhanulu ,Introduction to Lasers: theory and applications S.Chand publications 2012,New Delhi.



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**OBJECTIVES**

At the end of the course, the students should be able to:

- To make the students conversant with boiler feed water requirements, related problems and water treatment techniques.
- To recall the terminologies of electrochemistry and explain the function of batteries and fuel cells with its electrochemical reactions.
- To understand the fundamentals of corrosion, its types and polymers with its applications.
- Types of fuels, calorific value calculations, manufacture of solid, liquid and gaseous fuels

**UNIT I****WATER AND ITS TREATMENT**

9

Hardness of water - types - expression of hardness - units - estimation of hardness of water by EDTA - numerical problems - Alkalinity - types of alkalinity - determination of alkalinity - boiler troubles (scale and sludge) - treatment of boiler feed water - Internal treatment (carbonate, colloidal, phosphate and calgon conditioning) external treatment Ion exchange process, zeolite process - desalination of brackish water - Reverse Osmosis.

**UNIT II****ELECTROCHEMISTRY AND ENERGY STORAGE DEVICES**

9

Electrochemical cell - single electrode potential - standard electrode potential - electrochemical series and its significance - EMF of a cell - Nernst equation - Electrodes - Reference electrodes - hydrogen, calomel, quinhydrone and glass electrodes. Determination of pH of a solution using a glass electrode. Batteries - primary and secondary cells, dry cell, alkaline, lead acid storage cell, Ni-Cd battery and lithium nano battery. Clean energy fuel cells - H<sub>2</sub>-O<sub>2</sub> fuel cell.

**UNIT III****CORROSION SCIENCE**

9

Corrosion: definition - types of corrosion: chemical and electrochemical corrosion - Pilling Bedworth ratio - types of oxide layer (stable, unstable, volatile, porous) - hydrogen evolution and oxygen absorption mechanism for electrochemical corrosion - mechanism for rusting of iron. Types of electrochemical corrosion: Galvanic corrosion - differential aeration corrosion (pitting, waterline and pipeline). Galvanic series - applications. Factors influencing corrosion: nature of metal and environment. Corrosion control methods: sacrificial anode method - impressed current Cathodic protection method - electroplating - electroless plating.

**UNIT IV****POLYMERS AND ITS PROCESSING**

9

Monomers - polymers - polymerization - functionality - degree of polymerization - classification of polymers based on source and applications - Molecular weight determination. Types of polymerizations: addition, condensation and copolymerization - mechanism of free radical polymerization. Preparation, properties and applications of PE, PVC, Teflon, terylene, Nylon and Bakelite. Rubber - drawbacks of natural rubber - Vulcanization - Compounding of plastics - injection and blow moulding methods.

**UNIT V****FUELS AND COMBUSTION**

9

**Fuels:** Introduction - classification of fuels - coal - analysis of coal (proximate and ultimate) - carbonization - manufacture of metallurgical coke (Otto Hoffmann method) - petroleum - manufacture of synthetic petrol (Bergius process) - knocking - octane number - diesel oil -

cetane number - natural gas - compressed natural gas (CNG) - liquefied petroleum gases (LPG) - power alcohol and biodiesel. **Combustion of fuels:** Introduction - calorific value - higher and lower calorific values- theoretical calculation of calorific value - ignition temperature - spontaneous ignition temperature - explosive range - flue gas analysis (ORSAT Method).

**TOTAL HOURS 45 PERIODS**

### **COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

CO1: Attribute the internal and external treatment methods for the removal of hardness in water for domestic and industrial applications.

CO2: Construct an electrochemical cell and identify the components and processes in batteries and infer the selection criteria for commercial battery systems with respect to different applications.

CO3: Utilize electrochemical data to formulate an electrochemical half-cell and cell reactions for corrosion control processes.

CO4: Analyse the three types of fuels based on calorific value for selected application.


CO5: Analyse the three types of fuels based on calorific value for selected application.

### **TEXT BOOKS**

1. S. S. Dara and S. S. Umare, "A Textbook of Engineering Chemistry", S. Chand & Company LTD, New Delhi, 2015
2. P. C. Jain and Monika Jain, "Engineering Chemistry" Dhanpat Rai Publishing Company (P) LTD, New Delhi, 2015

### **REFERENCE BOOKS**

1. Friedrich Emich, "Engineering Chemistry", Scientific International PVT, LTD, New Delhi, 2014.
2. Prasanta Rath, "Engineering Chemistry", Cengage Learning India PVT, LTD, Delhi, 2015.
3. Shikha Agarwal, "Engineering Chemistry-Fundamentals and Applications", Cambridge University Press, Delhi, 2015.

  
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ENGINEERING GRAPHICS

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## OBJECTIVES

At the end of the course, the students should be able to:

- To understand the graphical skills for drawing the object and the principle of free-hand sketching techniques.
- To understand the principle of orthographic projection of points, lines and plane surfaces.
- To study the principle of simple solids.
- To understand the principle of section and development of solids.
- To understand the principle of Isometric and Perspective projections.

### Concepts and conventions (Not for Examination)

3

Importance of graphics in engineering applications – Use of drafting instruments – BIS conventions and

specifications – Size, layout and folding of drawing sheets – Lettering and dimensioning.

### UNIT I

#### PLANE CURVES AND FREE HAND SKETCHING

9+6

**Curves used in engineering practices:**

Conics – Construction of ellipse, Parabola and hyperbola by eccentricity method – Construction of cycloid – construction of involutes of square and circle – Drawing of tangents and normal to the above curves.

**Free hand sketching:**

Representation of Three-Dimensional objects – General principles of orthographic projection – Need for importance of multiple views and their placement – First angle projection – layout views – Developing visualization skills through free hand sketching of multiple views from pictorial views of objects.

### UNIT II

#### PROJECTION OF POINTS, LINES AND PLANE SURFACES

9+6

Projection of points and straight lines located in the first quadrant – Determination of true lengths and true inclinations – Projection of polygonal surface and circular lamina inclined to both reference planes.

### UNIT III

#### PROJECTION OF SOLIDS

9+6

Projection of simple solids like prisms, pyramids, cylinder and cone when the axis is inclined to one reference plane by change of position method.

### UNIT IV

#### SECTION OF SOLIDS AND DEVELOPMENT OF SURFACES

9+6

Sectioning of above solids in simple vertical position by cutting planes inclined to one reference plane and perpendicular to the other – Obtaining true shape of section. Development of lateral surfaces of simple and truncated solids – Prisms, pyramids, cylinders and cones – Development of lateral surfaces of solids with cylindrical cutouts, perpendicular to the axis.

### UNIT V

#### ISOMETRIC AND PERSPECTIVE PROJECTIONS

9+3

Principles of isometric projection – isometric scale – isometric projections of simple solids, truncated prisms, pyramids, cylinders and cones.



Perspective projection of prisms, pyramids and cylinders by visual ray method.

**Total Hours 45+30 PERIODS**

### **COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

CO1: Recognize the conventions and apply dimensioning concepts while drafting simple objects. CO2: Draw the orthographic projection of points, line, and plane surfaces.

CO3: Draw the orthographic projection of simple solids.

CO4: Draw the section of solid drawings and development of surfaces of the given objects.

CO5: Apply the concepts of isometric and perspective projection in engineering practice.

### **TEXT BOOKS**

1. Ranganath G, Channankaiah and Halesh Koti, "Engineering Graphics", Second Edition, Sahana Publishers, 2015.
2. Bhatt. N.D., "Engineering Drawing" Charotar Publishing House, 53<sup>th</sup> Edition, 2014.

### **REFERENCE BOOKS**

1. Dhananjay A. Jolhe, "Engineering Drawing with an introduction to AutoCAD" Tata McGraw Hill Publishing Company Limited, 2017.
2. Gopalakrishnana. K. R, "Engineering Drawing" (Vol. I & II), Subhas Publications, 2014.
3. Basant Agarwal and C.M. Agarwal, "Engineering Drawing", Tata McGraw Hill, 2013.

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**OBJECTIVES**

At the end of the course, the students should be able to:

- To gain the knowledge on civil works like masonry, roofing, flooring and plastering.
- To gain the knowledge on stress, strain of various building and foundations.
- The students should familiar with foundry, welding and forging processes.
- The students should familiarly work principle of IC engines and its types.
- To gain the knowledge about various energy recourses and refrigeration air conditions systems.

**UNIT I SURVEYING AND CIVIL ENGINEERING MATERIALS 9**

**Surveying:** Objects, types, classification, principles, measurements of distances, angles, leveling, determination of areas, illustrative examples. **Civil Engineering Materials:** Bricks, stones, sand, cement, concrete, steel sections.

**UNIT II BUILDING COMPONENTS AND STRUCTURES 10**

**Foundations:** Types, Bearing capacity, Requirement of good foundations. **Superstructure:** Brick masonry, stone masonry, beams, columns, lintels, roofing, flooring, plastering, Mechanics, Internal and external forces, Stress, Strain, Elasticity, Types of Bridges and Dams, Basics of Interior Design and Landscaping.

**UNIT III FOUNDRY WELDING AND FORGING 10**

**Foundry:** Introduction - Patterns –materials. Types of pattern and pattern allowances. Molding sand, types and properties, Molding procedure. **Welding:** Definition and Classification, Gas welding, Oxy Acetylene welding, Types of flames, advantages and disadvantages of gas welding. Resistance welding - Classification, Spot welding and Seam welding. Soldering, Definition and Classification. Brazing – Definition and Classification. **Forging:** Types of Forging, Differences between Hot working and Cold working processes.

**UNIT IV I C ENGINES & BOILERS 8**

Internal combustion engines, working principle of Petrol and Diesel Engines, Four stroke and Two stroke cycles, Comparison of four stroke and two stroke engines, Boilers: Introduction of boilers, classification, Lancashire boiler, Babcock and Wilcox boiler, list of boiler mountings and accessories and applications (no sketches).

**UNIT V SOURCE OF ENERGY & REFRIGERATION 8**

**Sources of energy:** Introduction, conventional and non-conventional sources of energy, examples, solar energy, hydro power plant. Introduction to refrigeration and air-conditioning, COP, properties of refrigerants and types of refrigerants, working principle of vapour compression & vapour absorption refrigeration system, Layout of typical domestic refrigerator, Window and Split type room Air conditioner.

**TOTAL HOURS 45 PERIODS**

**COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

CO1: The usage of surveying and properties of construction materials.

CO2: The stress strain of various building and material such as substructure, road transport

and bridge.

CO3: The concept of manufacturing methods encountered in engineering practice such as foundry, welding and forging processes.

CO4: The working of internal combustion engines and its types.

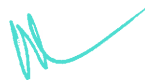
CO5: The concept of energy conservation in practical, power plant refrigeration air condition and its types.

#### **TEXT BOOKS**

1. Ranganath G and Channankaiah, “Basic Engineering Civil & Mechanical”, S.S.Publishers, 2014.
2. Shanmugam G and Palanichamy M S, “Basic Civil and Mechanical Engineering”, Tata McGraw Hill Publishing Co., New Delhi, 3rd Edition, 2012.

#### **REFERENCE BOOKS**

1. Venugopal.K and PrabhuRaja.V, “Basic Mechanical Engineering”, Anuradha Publishers, Kumbakonam, 2015.
2. Ramamrutham. S, “Basic Civil Engineering”, Dhanpat Rai Publishing Co. (P) Ltd, 3<sup>rd</sup> Edition reprint, 2013.
3. Shanmugasundaram. S and Mylsamy. K, “Basics of Civil and Mechanical Engineering”, Cenage Learning
4. India Pvt.Ltd, NewDelhi, 2012.
5. Khanna O.P, Foundry Technology, Dhanpat Rai Publishing Co. (P) Ltd, 2011.
6. Shanmugam G., “Basic Mechanical Engineering”, Tata McGraw Hill Publishing Co., New Delhi, 2010.



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**OBJECTIVES**

At the end of the course, the students should be able to:

- Students will be conversant with the estimation of various compound Bussing volumetric and instrumental analysis

**LIST OF EXPERIMENTS (A minimum of TEN experiments shall be offered)**

1. Estimation of Total hardness by EDTA
2. Determination of percentage of calcium in Lime Stone by EDTA
3. Estimation of chloride in water sample
4. Estimation of alkalinity of Water sample
5. Determination of DO in Water (Winkler's Method)
6. Determination of Rate of Corrosion of the given steel specimen by weight loss method (Without inhibitor)
7. Determination of Rate of Corrosion of the given steel specimen by weight loss method (With inhibitor)
8. Conduct metric titration (Simple acid base)
9. Conduct metric titration (Mixture of weak and strong acids)
10. Conduct metric titration using  $\text{BaCl}_2$  vs  $\text{Na}_2\text{SO}_4$
11. Potentiometric Titration ( $\text{Fe}^{2+}$  /  $\text{KMnO}_4$  or  $\text{K}_2\text{Cr}_2\text{O}_7$ )
12. PH titration (acid & base)
13. Determination of water of crystallization of a crystalline salt -Copper sulphate
14. Preparation of Bio Diesel by Trans etherification method

**TOTAL HOURS 45 PERIODS****COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

CO1: Carry out the volumetric experiments and improve the analytical skills.


CO2: Understand the maintenance and usage of analytical instruments and thereby develop their CO3: skills in the field of engineering.

CO4: Understand the principle and handling of electrochemical instruments and Spectrophotometer.

CO5: Apply their knowledge for protection of different metals from corrosion by using different inhibitors

**TEXT BOOKS**

1. Arthur I. Vogel's, "Quantitative Inorganic Analysis including Elementary Instrumental Analysis", ELBS, Group, 7th Edition, 2000.
2. Dr. K .Sivakumar, "Engineering Chemistry lab manual", S.S publishers, 2016.

  
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118EPP08

ENGINEERING PRACTICES LAB

L T P C

0 0 2 1

## OBJECTIVES

At the end of the course, the students should be able to:

- To get the knowledge on welding techniques and its types.
- To do the fitting operation on a given material. (Specimen)
- To carry out sheet metal operation.
- To know the principle involved in plumbing work.
- To do the carpentry work on a given work piece

## LIST OF EXPERIMENTS

### WELDING:

Study of electric Arc welding and Gas welding tools and equipments.

**Preparation of Arc welding and Gas welding models: i) Butt joint ii) Lap joint iii) T-joint.**

### FITTING:

Study of fitting tools and operations.

**Preparation of fitting models: i) V-fitting ii) Square fitting**

### SHEET METAL WORK:

Study of sheet metal tools and operations

**Preparation of sheet metal models: i) Tray ii) Funnel**

### PLUMBING WORKS:

Study of pipeline joints and house hold fittings.

**Preparation of plumbing models: Basic pipe connections with PVC and GI pipe fittings.**

### CARPENTRY:

Study of wooden joints and tools used in roofs, doors, windows, furniture.

**Preparation of carpentry models: i) Lap joint ii) Dovetail joint iii) T-Joint**

### DEMONSTRATION ON:

### ELECTRICAL ENGINEERING PRACTICE

Study of Electrical components and equipment's

Residential house wiring using switches, fuse, indicator, lamp and energy meter.

### ELECTRONICS ENGINEERING PRACTICE

Study of Electronic components –Resistor, color coding, capacitors etc

Soldering practice – components soldering in simple electric circuit & testing continuity

### COMPUTER HARDWARE AND SOFTWARE PRACTICE

Study of PC Hardware, Internet & World Wide Web and Productivity tools including Word, Excel, PowerPoint and Publisher.

**TOTAL HOURS 45 PERIODS**

## COURSE OUTCOMES

*Upon Completion of this course, students will be able to:*

CO1: Prepare simple Lap, Butt and T- joints using arc welding equipments.


- CO2: Prepare the rectangular trays and funnels by conducting sheet metal operation.
- CO3: Prepare the pipe connections and identify the various components used in plumbing.
- CO4: Prepare simple wooden joints using wood working tools.
- CO5: Demonstrate basic electrical, electronic and computer components based on their physical parameters and dimensions

#### **TEXT BOOKS**

1. Ranganath. G, & Channankaiah, "Engineering Practices Laboratory Manual" S.S. publishers, 2014.
2. Jeyapoovan.T &, S Gowri "Engineering Practice Lab Manual" Vikas publishing house Pvt.ltd, 2014.

#### **REFERENCE BOOKS**

1. Kannaiah.P & Narayana.K.L, "Manual on Workshop Practice", Scitech Publications, 2015.
2. Ramesh BabuV, "Engineering Practices Laboratory Manual", VRB Publishers Private Limited, Chennai,
3. Revised Edition, 2014.
4. Peter Norton, "Introduction to Computers", 7th Edition, Mc Graw Hill, 2010.
5. Bawa. H.S, "Workshop Practice", Tata McGraw – Hill Publishing Company Limited, 2009.
6. David Anfinson and Ken Quamme, "IT Essentials PC Hardware and Software Companion Guide", CISCO Press, Pearson Education, Third Edition, 2008.



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218ENT01

Communicative English

L T P C

3 0 2 3

## OBJECTIVES

At the end of the course, the students should be able to:

- To help learners develop their listening skills which will enable them listen to lectures and comprehend them by asking questions; seeking clarifications.
- To help learners develop their speaking skills and speak fluently in real contexts.
- To develop the basic reading and writing skills of first year engineering and technology students.
- To help learners develop grammar and vocabulary of a general kind by developing their reading skills

### UNIT I

9

Listening - conversation - Speaking – introducing oneself - exchanging personal information - Reading – comprehension - Writing - paragraph - Vocabulary Development - synonyms and antonyms - Language Development – consonants & vowels - phonetic transcription.

### UNIT II

9

Listening - telephonic conversation - Speaking – sharing information of a personal kind – greeting - taking leave - Reading – short stories – The Gift of the Magi, A Service of Love and The Last Leaf by O. Henry – Writing – developing hints - Vocabulary Development – everyday vocabulary - Language Development – British and American English - infinitive and gerund.

### UNIT III

9

Listening – class memory quiz - Speaking – impromptu - Reading – magazines – Writing – agenda - proposals - Vocabulary Development - important words used in speaking and writing - Language Development – types of sentences - information and emphasis.  
Agenda – Minutes of Meeting – Advertisement – Fliers – Brochures – Faxes – Internet Websites – Intranet Websites – Extranet Websites – Blog writing.

### UNIT IV

9

Listening – interviews of famous persons - Speaking – story narration - Reading – case study – Writing – invitation letter - quotation letter - Vocabulary Development – listening and reading vocabulary - Language Development – cause and effect – purpose and function.

### UNIT V

9

Listening - a scene from a film - Speaking - role play - Reading – jigsaw – Writing – essay writing - Vocabulary Development - business vocabulary - Language Development - degrees of comparison – real English phrases.

**TOTAL HOURS 45 PERIODS**

## COURSE OUTCOMES

*Upon Completion of this course, students will be able to:*

CO1: Comprehend conversations and talks delivered in English.

CO2: Participate effectively in formal and informal conversations; introduce themselves and

their friends and express opinions in English.

CO3: Read short stories, magazines, novels and other printed texts of a general kind.

CO4: Write short paragraphs, essays, letters and develop hints in English.

#### **TEXT BOOKS**

1. Board of Editors. Using English, A Coursebook for Undergraduate Engineers and Technologists. Orient BlackSwan Limited, Hyderabad: 2015.
2. Richards, C. Jack. Interchange Students' Book-2, New Delhi: CUP, 2015.
3. Uttham Kumar, N. Communicative English (with work book). Sahana Publications, Coimbatore, 2019.

#### **REFERENCE BOOKS**

1. Bailey, Stephen. Academic Writing: A Practical Guide for Students. New York: Rutledge, 2011.
2. Comfort, Jeremy, et al. Speaking Effectively: Developing Speaking Skills for Business English. Cambridge University Press, Cambridge: Reprint 2011.
3. Dutt P.Kiranmai and Rajeevan Geeta. Basic Communication Skills, Foundation Books:2013.
4. Means, L. Thomas and Elaine Langlois. English & Communication for Colleges. Cengage Learning, USA: 2007.
5. Redston, Chris & Gillies Cunningham. Face2Face (Pre-intermediate Student's Book & Workbook). Cambridge University Press, New Delhi: 2005.
6. Bailey, Stephen. Academic Writing: A Practical Guide for Students. New York: Rutledge, 2011.



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**Prerequisite** ENGINEERING MATHEMATICS-I

### OBJECTIVES

At the end of the course, the students should be able to:

- To understand double and triple integration concepts and apply to study vector calculus comprising of surface and volume integrals along with the classical theorems involving them.
- To learn analytic functions and their properties and also conformal mappings with few standard examples that have direct applications.
- To grasp the basics of complex integration and application to contour integration which is important for evaluation of certain integrals encountered in engineering problems.
- To introduce the concept of improper integrals through Beta and Gamma functions.

#### **UNIT I** **INTEGRAL CALCULUS** **9+3**

Definite and indefinite integrals - Substitution rule – Techniques of integration – Integration by parts – Trigonometric integrals - Trigonometric substitutions - Integration of rational functions by partial fractions – Integration irrational functions.

#### **UNIT II** **MULTIPLE INTEGRALS** **9+3**

Double integration – Cartesian and polar co-ordinates – Change of order of integration – Change of variables between Cartesian and polar coordinates – Triple integration in Cartesian co-ordinates – Area as double integral – Volume as triple integral.

#### **UNIT III** **VECTOR CALCULUS** **9+3**

Gradient Divergence and Curl – Directional derivative – Irrotational and solenoidal, vector fields – Vector integration – Green's theorem in a plane, Gauss divergence theorem and Stokes' theorem (Statement and applications only) – Simple applications involving cubes and rectangular parallelepipeds.

#### **UNIT IV** **ANALYTIC FUNCTIONS** **9+3**

Functions of a complex variable – Analytic functions – Necessary conditions, Cauchy–Riemann equation and Sufficient conditions (Statement and applications only) – Harmonic and orthogonal properties of analytic function (Statement and applications only) – Harmonic conjugate – Construction of analytic functions – Conformal mapping:  $w = z+c$ ,  $cz$ ,  $1/z$ , and bilinear transformation.

#### **UNIT V** **COMPLEX INTEGRATION** **9+3**

Complex integration – Statement and applications of Cauchy's integral theorem and Cauchy's integral formula – Taylor and Laurent expansions – Singular points – Residues – Residue theorem – Application of residue theorem to evaluate real integrals – Unit circle and semi-circular contour (excluding poles on boundaries).

**TOTAL HOURS 45 PERIODS**

### COURSE OUTCOMES

*Upon Completion of this course, students will be able to:*

CO1: Determine the area and volume in 2-dimension and 3-dimension respectively using

multiple integrals and also extending the concept to vector fields.

CO2: Learn the basic concepts of analytic functions and transformations of complex functions.

CO3: Master the integration in complex domain.


CO4: Understand the use of improper integrals' applications in the core subject.

#### **TEXT BOOKS**

1. Grewal. B.S., "Higher Engineering Mathematics", 43th Edition, Khanna Publications, Delhi, 2015.

#### **REFERENCE BOOKS**

1. James Stewart, "Stewart Calculus", 8th edition, 2015, ISBN: 9781285741550/1285741552.
2. Erwin Kreyszig, "Advanced Engineering Mathematics", tenth edition, Wiley India, 2011.
3. P.Kandasamy, K.Thilagavathy, K.Gunavathy, "Engineering Mathematics for first year", S.Chand & Company Ltd., 9th Edition, New Delhi, 2014.



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**OBJECTIVES**

At the end of the course, the students should be able to:

- To study the nature and facts about environment.
- To finding and implementing scientific, technological, economic and political solutions to environmental problems.
- To study the interrelationship between living organism and environment.
- To appreciate the importance of environment by assessing its impact on the human world;  
envison the surrounding environment, its functions and its value.
- To study the dynamic processes and understand the features of the earth's interior and surface.

**UNIT I****NATURAL RESOURCES**

14

Definition, scope and importance of environment – need for public awareness - Forest resources: Use and over- exploitation, deforestation, case studies- timber extraction, mining, dams and their effects on forests and tribal people – Water resources: Use and over-utilization of surface and ground water, floods, drought, conflicts over water, dams-benefits and problems – Mineral resources: Use and exploitation, environmental effects of extracting and using mineral resources, case studies – Food resources: World food problems, changes caused by agriculture and overgrazing, effects of modern agriculture, fertilizer-pesticide problems, water logging, salinity, case studies – Energy resources: Growing energy needs, renewable and non-renewable energy sources, use of alternate energy sources. case studies – Land resources: Land as a resource, land degradation, man induced landslides, soil erosion and desertification – role of an individual in conservation of natural resources – Equitable use of resources for sustainable lifestyles. Field study of local area to document environmental assets – river / forest / grassland / hill / mountain.

**UNIT II****ECOSYSTEMS AND BIODIVERSITY**

8

Concept of an ecosystem – structure and function of an ecosystem – producers, consumers and decomposers – energy flow in the ecosystem – ecological succession – food chains, food webs and ecological pyramids – Introduction, types, characteristic features, structure and function of the (a) forest ecosystem (b) grassland ecosystem (c) desert ecosystem (d) aquatic ecosystems (ponds, streams, lakes, rivers, oceans, estuaries) – Introduction to biodiversity definition: genetic, species and ecosystem diversity – biogeographical classification of India – value of biodiversity: consumptive use, productive use, social, ethical, aesthetic and option values – Biodiversity at global, national and local levels – India as a mega-diversity nation – hot-spots of biodiversity – threats to biodiversity: habitat loss, poaching of wildlife, man-wildlife conflicts – endangered and endemic species of India –conservation of biodiversity: In-situ and ex-situ conservation of biodiversity. Field study of common plants, insects, birds; Field study of simple ecosystems – pond, river, hill slopes.

**UNIT III****ENVIRONMENTAL POLLUTION**

10

Definition – causes, effects and control measures of: (a) Air pollution (b) Water pollution (c) Soil pollution (d) Marine pollution (e) Noise pollution (f) Thermal pollution (g) Nuclear

hazards – solid waste management: causes, effects and control measures of municipal solid wastes – role of an individual in prevention of pollution – pollution case studies – disaster management: floods, earthquake, cyclone and landslides. Field study of local polluted site – Urban / Rural / Industrial / Agricultural.

**UNIT IV SOCIAL ISSUES AND THE ENVIRONMENT 7**

From unsustainable to sustainable development – urban problems related to energy – water conservation, rain water harvesting, watershed management – resettlement and rehabilitation of people; its problems and concerns, case studies – role of non-governmental organization environmental ethics: Issues and possible solutions – climate change, global warming, acid rain, ozone layer depletion, nuclear accidents and holocaust, case studies. – wasteland reclamation – consumerism and waste products – environment production act – Air (Prevention and Control of Pollution) act – Water (Prevention and control of Pollution) act – Wildlife protection act – Forest conservation act – enforcement machinery involved in environmental legislation- central and state pollution control boards- Public awareness.

**UNIT V HUMAN POPULATION AND THE ENVIRONMENT 6**

Population growth, variation among nations – population explosion – family welfare programme – environment and human health – human rights – value education – HIV/AIDS – women and child welfare – role of information technology in environment and human health – Case studies.

**TOTAL HOURS 45 PERIODS**

**COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

CO1: Environmental Pollution or problems cannot be solved by mere laws. Public participation is an important aspect which serves the environmental Protection. One will obtain knowledge on the following after completing the course.


CO2: Public awareness of environmental is at infant stage.

CO3: Ignorance and incomplete knowledge has led to misconceptions

CO4: Development and improvement in std. of living has led to serious environmental disasters

**TEXT BOOKS**

1. Benny Joseph, Environmental Science and Engineering ‘, Tata McGraw-Hill, New Delhi, 2006.
2. Gilbert M. Masters, Introduction to Environmental Engineering and Science ‘, 2nd edition, Pearson Education, 2004.
3. Dr. G. Ranganath, Environmental Science and Engineering, Sahana Publishers, 2018 edition.

  
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**OBJECTIVES**

At the end of the course, the students should be able to:

- To understand the vectorial and scalar representation of forces and moments.
- To understand the static equilibrium of particles and rigid bodies both in two dimensions.
- To understand the concepts of centroids and moment of inertia of composite sections.
- To understand the principle of work and energy.
- To enable the students to comprehend the effect of friction on equilibrium

**UNIT I** **BASICS & STATICS OF PARTICLES** **9+3**  
 Introduction-Units and Dimensions-Laws of mechanics - Lame's theorem, Parallelogram and Triangular law of forces, Polygon force, Resolution and Composition of forces, Equilibrium of a particle-Forces in space - Equilibrium of a particle in space-Equivalent systems of forces-Principle of transmissibility-Single equivalent force.

**UNIT II** **EQUILIBRIUM OF RIGID BODIES** **9+3**  
 Free body diagram-Types of supports and their reactions-Requirements of stable equilibrium-Moments and Couples, Moment of a force about a point and about an axis-Vectorial representation of couples- Varignon's Theorem-Equilibrium of Rigid bodies in two dimensions-Equilibrium of Rigid bodies in three dimensions – Examples.

**UNIT III** **PROPERTIES OF SURFACES AND SOLIDS** **9+3**  
 Determination of Areas and Volumes-First moment of area and the centroid of sections - rectangle, circle, triangle from integration - T section, I section, Angle section, Hollow section by using standard formula, Second and product moments of plane area - Rectangle, triangle, circle from integration-T section, I section, Angle section, Hollow section by using standard formula, Parallel axis theorem and perpendicular axis theorem.

**UNIT IV** **DYNAMICS OF PARTICLES** **9+3**  
 Displacement, Velocity and Acceleration, their relationship, Relative motion- Rectilinear motion-Curvilinear motion, Newton's law-Work Energy Equation of particles-Impulse and Momentum-Impact of elastic bodies.

**UNIT V** **FRICITION** **9+3**  
 Frictional force - Laws of Coloumb friction - Simple contact friction - Rolling resistance - Belt friction - Ladderfriction - wedge friction.

**TOTAL HOURS 45+15 PERIODS**

**COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*


- CO1: Explain the differential principle applies to solve engineering problems dealing with force, displacement, velocity and acceleration.
- CO2: Find solution for problems related to equilibrium of particles.
- CO3: Solve the Moment of inertia for different 2-D plane figures.
- CO4: Analyze the forces in any structures.
- CO5: Solve rigid body subjected to frictional forces.

### **TEXT BOOKS**

1. Ramamrutham S, "Engineering Mechanics (S.I Units)", Dhanpat Rai Publications, 10th Edition, Reprint 2015.
2. Dr. Gujral I S, "Engineering Mechanics", Lakmi Publications, Second Edition, 2011.

### **REFERENCE BOOKS**

1. Bhavikatti S, "Engineering Mechanics", New Age International Publisher, 4th Edition, 2014.
2. Khurmi R S, "Engineering Mechanics", S Chand Publisher, 20th Edition, 2012.
3. Dr. Bansal R K and Sanjay Bansal, "Engineering Mechanics", Lakshmi Publication, 7th Edition, 2011.
4. Rajput R K, "Engineering Mechanics", Dhanpat Rai Publications, 3rd Edition, 2005.



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CO2: Read, write, execute by hand simple Python programs.

CO3: Structure simple Python programs for solving problems.

CO4: Decompose a Python program into functions.


CO5: Represent compound data using Python lists, tuples, dictionaries. CO6: Read and write data from/to files in Python Programs.

#### **TEXT BOOKS**

1. Allen B. Downey, ``Think Python: How to Think Like a Computer Scientist'', 2nd edition, Updated for Python 3, Shroff/O'Reilly Publishers, 2016 (<http://greenteapress.com/wp/think-python/>)
2. Guido van Rossum and Fred L. Drake Jr, —An Introduction to Python – Revised and updated for Python 3.2, Network Theory Ltd., 2011.

#### **REFERENCE BOOKS**

1. John V Guttag, —Introduction to Computation and Programming Using Python'', Revised and expanded Edition, MIT Press, 2013
2. Robert Sedgewick, Kevin Wayne, Robert Dondero, —Introduction to Programming in Python: An Inter-Disciplinary Approach, Pearson India Education Services Pvt. Ltd., 2016.
3. Timothy A. Budd, —Exploring Python, Mc-Graw Hill Education (India) Private Ltd., 2015.
4. Kenneth A. Lambert, —Fundamentals of Python: First Programs, CENGAGE Learning, 2012.

  
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## BIOCHEMISTRY

L T P C

318BTT02

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### OBJECTIVES

At the end of the course, the students should be able to:

- To learn the fundamentals of biochemical processes
- To learn the structure and properties of biomolecules and its function
- To gain knowledge of concepts of metabolism
- To gain knowledge of metabolic regulation and intermediate compounds
- To gain knowledge of transportation of protein and degradation

**UNIT I INTRODUCTION TO BIOMOLECULES-CARBOHYDRATES 9**  
Basic principles of organic chemistry, role of carbon, types of functional groups, chemical nature of water, pH and biological buffers, biomolecules. Structure and properties of Carbohydrates (mono, di, oligo & polysaccharides) Proteoglycans, glucosaminoglycans. mutarotation, glycosidic bond; reactions of monosaccharides, reducing sugars. Starch, glycogen, cellulose and chitin. Proteoglycans, glycosaminoglycans. hyaluronic acid, chondroitin sulfate.

**UNIT II STRUCTURE AND PROPERTIES OF OTHER BIOMOLECULES 9**  
Structure and properties of Important Biomolecules.  
**Lipids:** Fatty acids, glycerol, saponification, Iodination, hydrogenation, phospholipids, glycolipids, sphingolipids, cholesterol, steroids, prostaglandins.  
**Protein:** Amino Acids, Peptides, Proteins, measurement, structures, hierarchy of organization primary, secondary, tertiary and quaternary structures, glycoproteins, lipoproteins. Determine of primary structure.  
**Nucleic acids:** Purines, pyrimidines, nucleoside, nucleotide, RNA, DNA- Watson-Crick structure of DNA, reactions, properties, measurement, nucleoprotein complexes

**UNIT III CONCEPTS OF METABOLISM AND CARBOHYDRATE METABOLISM 9**  
Functions of Proteins, Enzymes, Introduction to biocatalysts, metabolic pathways, primary and secondary metabolites. Interconnection of pathways and metabolic regulation. Glycolysis, TCA cycle, gluconeogenesis, pentose phosphate shunt & glyoxalate shunt.

**UNIT IV INTERMEDIARY METABOLISM AND REGULATION 9**  
Fatty acid synthesis and oxidation, reactions of amino acids, deamination, transamination and decarboxylation, urea cycle, Bioenergetics-High energy compounds, electron negative potential of compounds, respiratory chain, ATP cycle, calculation of ATP yield during oxidation of glucose and fatty acids.

**UNIT V PROTEIN TRANSPORT AND DEGRADATION 9**  
Protein targeting, signal sequence, secretion; Folding, Chaperone and targeting of organelle proteins, Protein degradation, receptor-mediated endocytosis, turnover.

**TOTAL HOURS 45 PERIODS**

### COURSE OUTCOMES

*Upon Completion of this course, students will be able to:*

- CO1: To ensure students have a strong foundation in the structure and reactions of biomolecules.  
CO2: To understand metabolic pathways of the major biomolecules and relevance to clinical conditions.  
CO3: To correlate biochemical processes with biotechnology applications.  
CO4: To understand about metabolic regulation and intermediate compounds.  
CO5: To understand about protein secretion, folding, transportation and degradation.

### TEXT BOOKS


1. Lehninger Principles of Biochemistry 6th Edition by David L. Nelson, Michael M. Cox 2001
2. Satyanarayana, U. and U. Chakerapani, "Biochemistry" 3rd Rev. Edition, Books & Allied (P) Ltd., 2006. 31
3. Rastogi, S.C. "Biochemistry" 2nd Edition, Tata McGraw-Hill, 2003.
4. Conn, E.E., et al., "Outlines of Biochemistry" 5th Edition, John Wiley & Sons, 1987.
5. Outlines of biochemistry, 5th Edition: By E E Conn, P K Stumpf, G Bruening and R Y Doi. pp 693. John Wiley and Sons, New York. 1987.

### REFERENCE BOOKS

1. Berg, Jeremy M. et al. "Biochemistry", 6th Edition, W.H. Freeman & Co., 2006.
2. Murray, R.K., et al. "Harper's Illustrated Biochemistry", 27th Edition, McGraw-Hill, 2006.
3. Voet, D. and Voet, J.G., "Biochemistry", 3rd Edition, John Wiley & Sons Inc., 2004.

### EBOOKS/WEBLINKS

1. <http://dl4a.org/uploads/pdf/Biochemistry.pdf>
2. <http://www.louisbolk.org/downloads/1282.pdf>
3. <https://awesomechem.files.wordpress.com/2016/10/harpers-illustrated-biochemistry-28th-ed-robert-k-murray-et-al-mcgraw-hill-2009.pdf>

  
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**OBJECTIVES**

At the end of the course, the students should be able to:

- To provide knowledge on the fundamentals of cell biology
- To help students understand the signaling mechanisms
- To understand how organisms' function and the structure and functions of the plasma membrane and the major organelles that occur in prokaryotic and eukaryotic cells.
- To understand how cellular organelles work together to carryout life functions.
- To protect cells to prevent infection and other harmful effects.

**UNIT I CELL STRUCTURE AND FUNCTION**

9

Structure and function of Prokaryotic and Eukaryotic organelles, principles of membrane organization, membraneproteins, cytoskeletal proteins, types of cell division, mitosis & meiosis, extra cellular matrix, cell cycle and molecules that control cell cycle.

**UNIT II TRANSPORT ACROSS CELL MEMBRANES**

9

Passive & active transport, permeases, sodium potassium pump, Ca<sup>2+</sup> ATPase pumps, lysosomal and vacuolar membrane ATP dependent proton pumps, cotransport symport, antiport, transport into prokaryotic cells, endocytosis and exocytosis, Entry of viruses and toxins into cells

**UNIT III RECEPTORS AND MODELS OF EXTRACELLULAR SIGNALLING**

9

Cytosolic, nuclear and membrane bound receptors, Types of receptors and mode of action: autocrine, paracrine, endocrine, tyrosine kinases, G Protein receptor.

**UNIT IV SIGNAL TRANSDUCTION**

9

Signal amplification, different models of signal amplifications, cyclic amp, role of inositol phosphates as messengers, biosynthesis of inositol triphosphates, cyclic GMP and G proteins, role in signal transduction, calcium ion flux and its role in cell signaling, current models of signal amplification, MAP kinases, regulation of proteinkinases, serine –threonine kinases, tumor necrosis factor receptor families

**UNIT V CELL CULTURE**

9

Techniques for the propagation of eukaryotic cells. Cell lines, generation of cell lines, maintenance of stock cells, characterization of cells, morphological analysis techniques in cell culture, ex-plant cultures, primary cultures and differentiated cell line. *Cell fractionation and flow cytometry and Localization of proteins in cells– Immunostaining.*

**TOTAL HOURS 45 PERIODS**

**COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

CO1: To develop integral knowledge on cell structure, molecular organization and function of cell organelles.

CO2: To learn the cell is the basic unit of life in the entire living world.

CO3: To Understand the basic knowledge once structure and function as well as on the molecular basis of chromatin organization

CO4: Understand cell at structural and functional level.

CO5: Understand the molecular interaction between cells and signal transduction, secondary messengers.

**TEXT BOOKS**

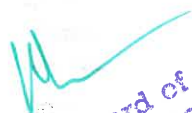
1. Molecular Cell Biology, Darnell J, Lodish H, Baltimore D W.H. Freeman 6<sup>TH</sup> Edition 2005.
2. Cell Biology Kimball T.W., Wesley Publishers, 3rd Edition, 2007.
3. The Cell Geoffrey Cooper, ASM Press, 2<sup>nd</sup> Edition 2007.
4. Molecular Biology of the Cell, James D. Watson, Wilkins, a Wolters Kluwer Business Publishers 8<sup>th</sup> Edition, 2013.

#### REFERENCE BOOKS

1. Cell Biology De Robertis & De Robertis, ASM Press and Sinauer Associates 4th Edition, 2000
2. Cell and Molecular Biology Ajoy Paul, Books and Allied (P) Ltd 2007.
3. Cell and Molecular Biology, Gerald Karp, Wiley Publishers, 7<sup>th</sup> Edition, 2013.

#### EBOOKS/WEBLINKS

1. <https://www.scribd.com/.../Karp-Cell-and-Molecular-Biology-Concepts>
2. <https://www.nyu.edu/projects/fitch/courses/moleccell/precellevo.pdf>
3. [http://web.iitd.ac.in/~amittal/SBL101\\_Essentials\\_Cell\\_Biology.pdf](http://web.iitd.ac.in/~amittal/SBL101_Essentials_Cell_Biology.pdf)
4. [https://edisciplinas.usp.br/pluginfile.php/86323/mod\\_resource/content/1/MolecularBiologyOfTheCell5th.Ed-pag579+37.pdf](https://edisciplinas.usp.br/pluginfile.php/86323/mod_resource/content/1/MolecularBiologyOfTheCell5th.Ed-pag579+37.pdf)

  
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**OBJECTIVES**

At the end of the course, the students should be able to:

- To introduce students to the principles of Microbiology to emphasize structure and biochemical aspects of various microbes
- To enable students, learn the basic structure, growth and metabolism of microorganisms
- To solve the problems in microbial infection and their control
- To enable students to learn the production process and preservation techniques
- To develop skills of the students in the area of industrial and environmental microbiology

**UNIT I BASIC TOOLS AND TECHNIQUES 9**

History of microbiology, classification and nomenclature of microorganisms, microscopic examination of microorganisms, light and electron microscopy, different staining techniques: gram staining, acid fast, capsular staining, flagellar staining.

**UNIT II MICROBES-STRUCTURE AND MULTIPLICATION 9**

Structural organization and multiplication of bacteria, viruses, algae and fungi, with special mention of life cycle history of actinomycetes, yeast, mycoplasma and bacteriophages.

**UNIT III MICROBIAL NUTRITION, GROWTH AND METABOLISM 9**

Nutritional requirements of bacteria; different media used for bacterial culture; growth curve and different methods to quantify bacterial growth; aerobic and anaerobic bioenergetics and utilization of energy for biosynthesis of important molecules, biological control of microorganism.

**UNIT IV CONTROL OF MICROORGANISMS 9**

Physical and chemical control of microorganisms; host-microbe interactions; anti-bacterial, antifungal and anti-viral agents; mode of action and resistance to antibiotics; clinically important microorganisms- *Bacillus subtilis*, *Clostridium botulinum*.

**UNIT V INDUSTRIAL AND ENVIRONMENTAL MICROBIOLOGY 9**

Primary metabolites; secondary metabolites and their applications; preservation of food; *broad spectrum antibiotics*, production of penicillin, alcohol, vitamin B-12; biogas; bioremediation; leaching of ores by microorganisms; biofertilizers and biopesticides; microorganisms and pollution control.

**TOTAL HOURS 45 PERIODS**

**COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

- CO1: Students attains knowledge on the principles of Microbiology and biochemical aspects of various microbes
- CO2: Knowledge on the microorganisms Structure and its different types, growth and metabolism
- CO3: The interactions between contaminants, soil, water and microorganisms and its control
- CO4: Knowledge on the production process and preservation techniques
- CO5: An ability to conduct experiments, as well as to analyze and interpret data

**TEXT BOOKS**

1. Prescott L. M., Harley J. P., Klein DA, Microbiology, 3<sup>rd</sup> Edition, Wm. C. Brown Publishers, 1996.
2. Pelczar MJ, Chan ECS and Krein NR, Microbiology, Tata McGraw Hill Edition, New Delhi, India. 2005
3. Cruiger, Wulf and Anneliese Cruieger, "Biotechnology: A Text book of Industrial Microbiology",


IIInd Edition, Panima Publishing,2000.

#### REFERENCE BOOKS

1. Talaron K, Talaron A, Casita, Pelczar and Reid. Foundations in Microbiology, W. C. Brown Publishers,1993.
2. Casida, L. E. "Industrial Microbiology", New Age International(P)Ltd,1968
3. Stanier, RY., *et.al.*, General Microbiology, 5<sup>th</sup> ed. MacmillanPress.2000

#### EBOOKS/WEBLINKS

1. <https://nptel.ac.in/courses/102103015>
2. <https://openstax.org/details/books/microbiology>
3. Atlas,RM., Principles of Microbiology, 2nded.,1997,McGraw-Hill
4. <http://www.wwnorton.com/college/biology/microbiology2/>



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**OBJECTIVES**

At the end of the course, the students should be able to:

- To gain knowledge on basics of measurement
- To have a fundamental knowledge about the Light spectrum and Absorption.
- To understand working principles of Fluorescence NMR, Mass spectroscopy
- To acquire knowledge on the different chromatographic methods for separation of biological products and surface microscopy
- To gain knowledge about separation of biological products.

**UNIT I BASICS OF MEASUREMENT 9**

Classification of methods–types of noise–calibration of instrumental methods–electrical components and circuits – signal to noise ratio– signal– noise enhancement.

**UNIT II OPTICAL AND THERMAL METHODS 9**

General design–sources of radiation–wave lengths selectors–sample containers–radiation transducers – types of optical instruments–Calorimeter, Fluorimeter, Nephelometry– Fourier transform measurements. Thermo- gravimetric methods – differential thermal analysis–differential scanning calorimetry. Isothermal titration calorimetry.

**UNIT III MOLECULAR SPECTROSCOPY 9**

Measurement of transmittance and absorbance – Lambert Beer's law – spectrophotometer analysis –qualitative and quantitative absorption measurements - types of spectrometers – UV–visible – IR–Raman spectroscopy, NMR, ESR, SPR, MS–instrumentation – theory.

**UNIT IV ELECTRO ANALYSIS AND SURFACE MICROSCOPY 9**

Electrochemical cells- Electrode potential cell potentials – potentiometry- reference electrode – ion selective and molecular selective electrodes – Instrument for potentiometric studies – Voltametry – Cyclic and pulse voltametry- Applications of voltametry. Study of surfaces–Scanning probe microscopes– AFM and STM.

**UNIT V SEPARATION METHODS 9**

Introduction to chromatography – van Deemter equation–Thin Layer Chromatography) Paper Chromatography- gas chromatography–stationary phases–detectors–HPLC–pumps–columns–detectors – ion exchange chromatography– size exclusion chromatography– Agarose Electrophoresis, capillary electrophoresis-Adsorption Chromatography.

**TOTAL HOURS 45 PERIODS**

**COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

CO1: Understand and apply the concept of optical and thermal methods

CO2: Understand spectroscopy.

CO3: Understand principle of surface microscopy and its application

CO4: Acquire knowledge on separation techniques used for biological products

CO5: Acquire knowledge on different chromatographic methods for separation of biological products

**TEXT BOOKS**

1. Instrumental Methods of Analysis; Willard & H.Merrit, Phi, 1997thEdition CBSPublishers.
2. Instrumental Methods of Analysis, D. Skoog, 2000 5thEdition CollegePublishers.
3. Instrumental Methods of Chemical Analysis Galen N. Ewing 5th Edition McGraw Hill International 2006.

## REFERENCE BOOKS

1. Introduction to Instrumental Analysis by Robert D Braun, Pharma Book Syndicate 2005.
2. Instrumental Methods of Chemical Analysis by H Kaur PPM Publishers 1999.
3. Biophysical Chemistry by Upadhyay 4<sup>th</sup> Edition by Himalaya Publishing House 2007.

## EBOOKS/WEBLINKS

1. <http://web.unizlovdv.bg/plamenpenchev/mag/books/anchem/Handbook.pdf>
2. <https://marianoshraderkels.files.wordpress.com/2017/05/instrumental-methods-of-analysis-oxford-higher-education-by-sivasankar.pdf>
3. <https://marianoshraderkels.files.wordpress.com/2017/05/instrumental-methods-of-analysis-oxford-higher-education-by-sivasankar.pdf>



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**OBJECTIVES**

At the end of the course, the students should be able to:

- To make the students aware of the overall industrial bioprocesses has to help them to manipulate the process to the requirement of the industrial needs.
- The course prepares the students for the bulk production of commercially important modern bioproducts.
- To understand the production and purification of industrial enzymes,
- To gain knowledge about products of plant, animal and fungal cell cultures.
- To understand the production and purification of therapeutic proteins

**UNIT I INTRODUCTION TO INDUSTRIAL BIOPROCESSES 9**

A historical overview of industrial fermentation process, Definition and scope of Industrial Biotechnology, Stock culture, A brief survey of organisms, processes. Growth curve of microorganisms (Bacteria), Process flow sheeting –block diagrams, pictorial representation.

**UNIT II PRODUCTION OF PRIMARY METABOLITES**

A brief outline of processes for the production of some commercially important organic acids (e.g.citric acid, lactic acid, acetic acid); amino acids (glutamic acid, aspartic acid ) and alcohols (ethanol, butanol)

**UNIT III PRODUCTION OF SECONDARY METABOLITES 9**

Study of production processes for various classes of secondary metabolites: antibiotics: betalactams (penicillin, cephalosporin), aminoglycosides (streptomycin etc) macrolides (erythromycin), vitamins andsteroids.

**UNIT IV PRODUCTION OF ENZYMES AND OTHER BIOPRODUCTS 9**

Production of industrial enzymes such as proteases, amylases, lipases, cellulases. Production of biopesticides, biofertilisers, biopreservatives (Nisin), cheese, biopolymers (PHB), single cell protein.

**UNIT V PRODUCTION MODERN BIOTECHNOLOGY PRODUCTS 9**

Production of recombinant proteins and diagnostic applications, production of vaccines. Production of monoclonal antibodies.

**TOTAL HOURS 45 PERIODS**

**COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

CO1: Understand of the steps involved in the production of bioproducts

CO2: Understand the basic biotechnological engineering principles and models to do tasks

CO 3: Understand the Design and deliver useful modern biotechnology products to the society.

CO4: Understand the bulk production of commercially important modern bioproducts.

CO5: Understand the production and purification of Industrial Enzymes and products of plant and animal cell cultures.

**TEXT BOOKS**

1. Satyanarayana,U. "Biotechnology"Books&Allied(P)Ltd.,2005
2. Kumar, H.D. "A Textbook on Biotechnology" IInd Edition. Affiliated East West Press Pvt.Ltd.,1998.
3. Balasubramanian,D. etal.,"ConceptsinBiotechnology"UniversitiesPressPvt.Ltd.,2004.
4. Presscott, S.C.andCecilG.Dunn,"Industrial Microbiology",Agrobios(India),2005.
5. Dubey, R.C. "AText book of Biotechnology" S. Chand & Co.Ltd.,2006.

**REFERENCE BOOKS**

1. Casida, L.E. "Industrial Microbiology", New Age International(P)Ltd,1968.
2. Cruger, Wulf and Anneliese Crueger, "Biotechnology: A Text book of Industrial Microbiology", IInd Edition, Panim a Publishing, 2000.
3. Stanbury, P.F., A. Whitaker and S.J. Hall "Principles of Fermentation Technology", II<sup>nd</sup> Edition, Butterworth– Heinemann (an imprint of Elsevier), 1995.

#### **EBOOKS/WEBLINKS**

1. <http://www.thanut-swu.com/images/BOT101/BiotechnologyBook.pdf>
2. [http://www.absinitiative.info/fileadmin/media/Knowledge\\_Center/Pulications/Sectoral\\_Briefs/Sectoral\\_Brief](http://www.absinitiative.info/fileadmin/media/Knowledge_Center/Pulications/Sectoral_Briefs/Sectoral_Brief)
3. [- Biotech - 2015.pdf](#)
4. <https://www.pdfdrive.com/biology-and-biotechnology-e22686316.html>

  
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**OBJECTIVES**

At the end of the course, the students should be able to:

- To understand the principle of qualitative analysis of various biomolecules.
- To understand the concept of quantitative estimation of biomolecules.
- To understand the preparation of standard buffer solution
- To understand the analysis of the body fluids.
- To understand a strong foundation in the structure and reactions of Biomolecules

**LIST OF EXPERIMENTS**

1. Preparation of buffers and measurement of weak acid, base.
2. Qualitative analysis of carbohydrates (monosaccharides, disaccharides, polysaccharides etc.).
3. Qualitative analysis of proteins and amino acids.
4. Qualitative analysis of lipids (triglycerides, cholesterol, phospholipids etc.).
5. Quantitative analysis of carbohydrates (Benedict's method etc.)
6. Quantitative estimation of blood glucose (Anthrone Method)
7. Protein estimation by Lowry's method.
8. Protein estimation by Biuret method.
9. Quantitative estimation of amino acids by Ninhydrin method.
10. Estimation of DNA by Diphenylamine method.
11. Estimation of RNA by Orcinol method
12. Extraction of lipids and analysis by TLC
13. Enzymatic assay of phosphates.
14. Enzymatic hydrolysis of starch.

**TOTAL HOURS 45 PERIODS**

**COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

- CO1: Learning the principles behind the qualitative estimation of biomolecules.  
CO2: Understanding the principles behind quantitative estimation of biomolecules.  
CO3: Understanding the analysis of the same in the body fluids professional career  
CO4: Understanding the preparation of standard buffer solution.  
CO5: To ensure students have a strong foundation in the structure and reactions of Biomolecules.

**TEXT BOOKS**

1. Lehninger Principles of Biochemistry 6th Edition by David L. Nelson, Michael M. Cox
2. Satyanarayana, U. and U. Chakerapani, "Biochemistry" 3rd Rev. Edition, Books & Allied (P) Ltd., 2006. 31
3. Rastogi, S.C. "Biochemistry" 2nd Edition, Tata McGraw-Hill, 2003.

**REFERENCE BOOKS**

1. Wilson and Walker "Principles and Techniques of Practical Biochemistry" 4 Edn., Cambridge Knew pros 1997.
2. Plummer D T "An Introduction to Practical Biochemistry" III Edn., Tata McGrawhill.
3. Voet, D. and Voet, J. G., "Biochemistry", 3<sup>rd</sup> Edition, John Wiley & Sons Inc., 2004

**EBOOKS/WEBLINKS**

1. <http://www.louisbolk.org/downloads/1282.pdf>
2. <https://awesomechem.files.wordpress.com/2016/10/harpers-illustrated-biochemistry-28th-ed-robert-k->
3. <murray-et-al-mcgraw-hill-2009.pdf>



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**OBJECTIVES**

At the end of the course, the students should be able to:

- To learn the morphology, identification and propagation of cells
- To understand the basic techniques to work with cells
- To understanding and perform cell staining techniques
- To learn working principles of Microscopy
- To understand isolation of plasmids, nucleus or other organelles and cell division.

**LIST OF EXPERIMENTS**

1. Sterilization techniques.
2. Identification of plant, animal and their components by microscopy.
3. Isolation of chloroplast
4. Isolation of DNA from cauliflower
5. Determination of cell mobility-Hanging Drop method
6. Tryphan Blue Assay
7. Lactophenol Cotton Blue Staining
8. Osmosis and Tonicity.
9. Simple Staining.
10. Propagation and Maintenance of Cells
11. Staining for different stages of mitosis in *Allium Cepa* (Onion).

**TOTAL 45 PERIODS****COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

CO1: To learn the basic skills in light microscopy, cell fractionation, and spectroscopy.

CO2: To be able to perform light microscopy techniques, to isolate plastids, nucleus or other organelles and cell division.

CO3: To be able to identify the various stages of mitosis.

CO4: To understand the basic techniques to work with cells

CO5: To understand and perform cell staining techniques

**TEXT BOOKS**

1. "Laboratory Investigations in Cell and Molecular Biology", Allen Bregman Wiley publishers, 4<sup>th</sup> Edition, 2001.
2. "General Microbiology" Powar and Dagainawala, Himalaya Publishing House, 8<sup>th</sup> edition 2012.
3. "Cell Biology: A Laboratory Hand book Volume", Julio E. Celis, Tony Hunter Elsevier Academic Press, 3<sup>rd</sup> Edition, 2006.

**REFERENCE BOOKS**

1. "Cell Biology: A Laboratory Handbook: 004", Julio E. Celis, Academic Pr; 2<sup>nd</sup> edition, 3<sup>rd</sup> Edition, 2005.
2. "Laboratory Exercises and Techniques in Cellular Biology", Anthony Contento, Wiley Publishers, 1<sup>st</sup> Edition 2012
3. "Laboratory Methods in Cell Biology" S.Jha Academic Press, 1<sup>st</sup> Edition, 2012.

**EBOOKS/WEBLINKS**

1. <https://www.scribd.com/.../Karp-Cell-and-Molecular-Biology-Concepts>
2. <https://www.nyu.edu/projects/fitch/courses/moleccell/precellevo.pdf>
3. [http://web.iitd.ac.in/~amittal/SBL101\\_Essentials\\_Cell\\_Biology.pdf](http://web.iitd.ac.in/~amittal/SBL101_Essentials_Cell_Biology.pdf)

**OBJECTIVES**

At the end of the course, the students should be able to:

- The course aims to develop the skills of students in different areas of microbiology
- To demonstrate various techniques to learn the morphology, identification and propagation of microbes
- To solve the problems in microbial infection and their control
- To enable students, learn the basic structure, growth and metabolism of microorganisms
- To demonstrate various techniques on effect of physical Factors

**LIST OF EXPERIMENTS**

1. Introduction, Laboratory Safety, Use of Equipment; Sterilization Techniques
2. Culture Media- Types and Use; Preparation of Nutrient broth and agar
3. Culture Techniques, Isolation and Preservation of Cultures-Broth: flask, testtubes; Solid: Pourplates, streak plates, slants, stabs
4. Microscopy- Working and care of Microscope
5. Microscopic Methods in the Study of Microorganisms., Microscopic identification of yeast/ mould
6. Staining Techniques Simple, Differential-Gram's Staining, spore/capsule staining
7. Quantification of Microbes: Sampling and Serial Dilution; Bacterial count in Soil-TVC
8. Effect of Disinfectants-Phenol Coefficient
9. Antibiotic Sensitivity Assay
10. Growth Curve in Bacteria and Yeast
11. Effect of pH, Temperature, UV radiation on Growth Bacteria

**TOTAL HOURS 45 PERIODS****COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

CO1: Understand the advanced technical information pertaining to laboratory bio-safety and preventive measures from pathogenic microorganism.

CO2: Know the various aseptic techniques and sterilization methods

CO3: Understand the interactions between contaminants, soil, water and microorganisms and its control.

CO4: Gain knowledge on the microorganism structure and its different types, growth and metabolism

CO5: Develop the Skills to work on several important techniques for the study of microorganisms in the laboratory.

**TEXT BOOKS**

1. Cappuccino, J.G. and N.Sherman "Microbiology: A Laboratory Manual",4<sup>th</sup> Edition, Addison-Wesley,1999.
2. Collee, J.G. et al., "Mackie & McCartney Practical Medical Microbiology"4<sup>th</sup> Edition, Churchill Livingstone,1996.
3. Powarand dagainawala, "General microbiology", Himalaya Publishing House,2nded.2011

**REFERENCE BOOKS**

1. Salle, A.J., Fundamental Principles of Bacteriology,7<sup>th</sup>ed.,1999, Tata-McGrawHill,1998
2. Dubey, R.C., and Maheswari, D.K. Textbook of Microbiology, S.Chand & Co.2006
3. SubbaRao, NS. Soil Microbiology,4<sup>th</sup> Ed., Oxford & IBH Publishing Co. Pvt. Ltd.2018

**OBJECTIVES**

At the end of the course, the students should be able to:

- To impart the knowledge of basic probabilistic theory.

To learn one dimensional discrete and continuous probability distributions occurring in natural phenomena.

To extend the probability theory to two-dimensional random variable and to study the statistical measures.

To introduce the notion of sampling distributions and acquire the knowledge of statistical techniques useful in decision making.

To expose the statistical methods for analysis of variance and control limits.

**UNIT I PROBABILITY AND RANDOM VARIABLES 9+3**

Axioms of probability - Conditional probability - Total probability - Baye's theorem- Random variables - Probability mass function - Probability density function - Properties - Moments - Moment generating functions and their properties.

**UNIT II PROBABILITY DISTRIBUTIONS 9+3**

Binomial, Poisson, Geometric, Uniform, Exponential, and Normal distributions and their properties - Functions of a random variable-simple applications.

**UNIT III TWO-DIMENSIONAL RANDOM VARIABLES 9+3**

Joint distributions - Marginal and Conditional distributions - Covariance - Correlation and Linear regression - Central limit theorem (Statement and applications only for independent and identically distributed random variables).

**UNIT IV TESTING OF HYPOTHESIS 9+3**

Sampling distributions - Tests for single mean, Proportion, Difference of means (large and small samples) - Tests for single variance and equality of variances - Chi-square test for goodness of fit - Independence of attributes.

**UNIT V DESIGN OF EXPERIMENTS 9+3**

Analysis of variance - Completely Randomized Design (CRD) (one way classification) - Randomized Block Design (RBD) (two way classification) - Latin Square Design (LSD) - Factorial Designs-  $2^2$  factorial designs- Control charts for measurements -  $\bar{x}$  chart, R-chart,  $p$  - chart and  $np$  - chart.

**TOTAL HOURS 45 PERIODS****COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

CO1: To impart the knowledge of basic probabilistic theory.

CO2: To learn one dimensional discrete and continuous probability distributions occurring in natural phenomena.

CO3: To extend the probability theory to two-dimensional random variable and to study the statistical measures.

CO4: To introduce the notion of sampling distributions and acquire the knowledge of statistical

techniques useful in decision making.


CO5: To expose the statistical methods for analysis of variance and control limits.

**TEXT BOOKS**

1. Miller and Freund., "Probability and Statistics for Engineers", Pearson Education, Asia, 7th edition, 2012.
2. Veerarajan.T., "Probability, Statistics and Random Processes", Tata McGraw-Hill publishing company Limited, New Delhi, 2014

**REFERENCE BOOKS**

1. Spiegel, M.R, Schiller, J and Alu Srinivasan, R, "Schaum's Outlines Probability and Statistics", Tata McGraw-Hill Publishing Company Ltd. New Delhi, 2010.
2. Gupta.S.C., & Kapoor,V.K., "Fundamentals of mathematical statistics", 11th edition, Sultan Chand & Sons publishers, New Delhi, 2013.
3. Ibe, O.C., "Fundamentals of Applied Probability and Random Processes", Elsevier, U.P., 1<sup>st</sup> Indian Reprint, 2007.
4. Kandasamy. P, Thilagavathy, K., & Gunavathi. K., "Probability, Statistics and Queueing Theory", S. Chand & Company Ltd., New Delhi, 2014.
5. Hwei Hsu, "Schaum's Outline of Theory and Problems of Probability, Random Variables and Random Processes", Tata McGraw Hill edition, New Delhi, 2014.

  
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**OBJECTIVES**

At the end of the course, the students should be able to:

- Study the structural and functional organization of nucleic acids
- Learn molecular tools for studying activity of genes
- Learn the structure and properties of biomolecules and their functions
- Understand the genetics of prokaryotes and eukaryotes
- Acquire basic fundamental knowledge and explore skills in molecular biology and become aware of the complexity and harmony of cells.

**UNIT I CHEMISTRY OF NUCLEIC ACIDS**

9

Overview of Central dogma. Organization of prokaryotic and eukaryotic genome. Introduction to nucleic acids: Nucleic acids as genetic material, Structure and physicochemical properties of DNA and RNA elements, biological significance of differences in DNA and RNA. Primary structure of DNA: Chemical and structural qualities of 3',5'-Phosphodiester bond. Secondary Structure of DNA: DNA supercoiling. Reversible denaturation and hyperchromic effect.

**UNIT II DNA REPLICATION & REPAIR**

9

DNA replication: Meselson & Stahl experiment, bi-directional DNA replication, Okazaki fragments, Proteomics of DNA replication, Inhibitors of DNA replication, Overview of differences in prokaryotic and eukaryotic DNA replication, D-loop and rolling circle mode of replication. DNA mutations and their mechanism, various types of repair mechanisms.

**UNIT III TRANSCRIPTION**

9

Structure and function of mRNA, rRNA and tRNA. Structural aspects of gene. RNA synthesis, Proteins of RNA synthesis, Fidelity of RNA synthesis, Inhibitors of transcription, Differences in prokaryotic and eukaryotic transcription. Basic concepts in RNA world: Ribozymes, RNA processing and RNA editing.

**UNIT IV TRANSLATION**

9

Introduction to Genetic code: Elucidation of genetic code, Codon degeneracy, Wobble hypothesis and its importance, Prokaryotic and eukaryotic ribosome. Steps in translation: Initiation, Elongation and termination of protein synthesis prokaryotic and eukaryotic. Post translational modifications and their significance.

**UNIT V REGULATION OF GENE EXPRESSION**

9

Hierarchical levels of gene regulation, Prokaryotic gene regulation –*lac* and *trp* operon, Regulation Of gene expression with reference to  $\lambda$  phage life cycle.

**TOTAL HOURS 45 PERIODS**

**COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

- CO1: Ensure Have the basic knowledge of structure and biochemistry of nucleic acids and proteins and discriminate between them
- CO2: Understand the principles of DNA replication, transcription and translation and explain how they relate to each other
- CO3: Correlate Biochemical processes with molecular biology applications
- CO4: Understand metabolic regulation and intermediate compounds
- CO5: Understand gene organization and mechanisms of control of the gene and expression in various organisms

**TEXT BOOKS**


1. Friefelder, David. "Molecular Biology." Narosa Publications, 1999
2. Weaver, Robert F. "Molecular Biology" II<sup>nd</sup> Edition, Tata McGraw-Hill, 2003.
3. Karp, Gerald "Cell and Molecular Biology: Concepts and Experiments" IV<sup>th</sup> Edition, John Wiley, 2005.
4. Friefelder, David and George M. Malacinski "Essentials of Molecular Biology" II<sup>nd</sup> Edition, Panima Publishing, 1993.
5. Phundan Singh, "Principles of Genetics", Kalyani Publishers, 2012.

#### REFERENCE BOOKS

1. Dr. P. K. Gupta, "A Text Book of Cell & Molecular Biology" 4<sup>th</sup> Revised Edition, Rastogi Publications, 2015.
2. Robert Brooker, "Genetics: Analysis and Principles" 5<sup>th</sup> Edition, Publishing Pennsylvania Plaza publisher, 2014,
3. Dr. P. S. Verma and V K Agarwal, "Genetics", S. Chand publishing, 2010.

#### EBOOKS/WEBLINKS

1. <https://pothi.com/pothi/book/ebook-kaushlendraipathi-introduction-molecular-biology>
2. <https://www.kobo.com/us/en/ebook/biochemistry-and-molecular-biology>
3. <http://www.digitalbookindex.org/search/search010biolmolecularcellbiologya.asp>

  
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418BTT03	STOICHIOMETRIC AND PROCESS CALCULATIONS	L	T	P	C
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**Prerequisite** Engineering Mathematics II

## OBJECTIVES

At the end of the course, the students should be able to:

- To learn the basic principles of process calculations
- To understand the calculations of mass flow rate in different processes employed in bio-chemical industries
- To predict the energy consumption and energy efficiency in chemical processing industries
- To develop skills in the area of chemical engineering with emphasis on fluid mechanics
- To study the techniques and skills underlying fluid flow measurement.

### UNIT I BASIC PRINCIPLES OF MATERIAL BALANCES AND ENERGY BALANCES 9+3

Importance of material balance and energy balance in process industry Dimensions, Units, Conversion factors and their uses; applied mathematics for experimental curve fitting; Numerical differentiation; Numerical Integration

### UNIT II MATERIAL BALANCES

9+3

*Basic concepts involved in material balance calculations* - Overall and component balances; material balances without chemical reaction; material balances with chemical reactions *stoichiometric equation*, stoichiometric coefficient, *stoichiometric ratio*, *stoichiometric proportion*; degrees of freedom; recycle ratio calculations, purge ratio calculations; humidity calculations

### UNIT III ENERGY BALANCES

9+3

Overall and component balances; Calculation of heat capacity, specific heat capacity; partial pressure-calculations; Latent heats – calculations, energy balances - calculations, *Heat of mixing*, Sensible heat calculations; vapour pressure - calculations

### UNIT IV FLUID MECHANICS

9+3

Fluid-properties–*Fluid flow phenomena*–compressible, incompressible fluids, Newtonian And Non-Newtonian Fluids, Fluid statics for compressible & incompressible fluids applications in chemical engineering, Fluid pressure drop calculations. Pressure measuring devices

### UNIT V FLOW THROUGH PACKINGS AND FLUIDIZATION

9+3

Flow Measurement Orifice Meter, Venturimeter, Pitot tube; Flow in packed columns, flow in fluidization columns, settling phenomena- sedimentation, centrifugal pumps, centripetal pumps and *Reciprocating pumps*–characteristics, working and its applications

**TOTAL HOURS 45 PERIODS**

## COURSE OUTCOMES

*Upon Completion of this course, students will be able to:*

- CO1: Solve problems related to units and conversions and fit the given data using different methodologies
- CO2: Solve problems related to material balance concepts & design reactors for biochemical processes
- CO3: Solve problems related to energy balance concepts & design reactors for biochemical processes
- CO4: Apply their knowledge in describing the physical properties of fluid and calculating the pressure distribution for incompressible fluids and
- CO5: Design a system, component, or process to meet desired needs within realistic constraints such as economic, manufacturability, and sustainability.

### TEXT BOOKS


1. McCabe, W.L., J.C. Smith and P. Harriot "Unit Operations of Chemical Engineering", 6<sup>th</sup> Edition, McGrawHill, 2014.
2. Bhatt, B.I. and S.M. Vora "Stoichiometry (SI Units)", 3<sup>rd</sup> Edition, Tata McGraw-Hill, 2014.
3. K.A. Gavhane, "Introduction to process calculations", 2<sup>nd</sup> Edition, Nirali Prakashan 2012.
4. Narayanan, K.V. and Lakshmi Kutty "Stoichiometry and Process Calculations", 2<sup>nd</sup> Edition, PHI, 2006.
5. Geankoplis, C. J. "Transport Processes and Separation Process Principles", 7<sup>th</sup> Edition, PHI, 2012.

### REFERENCE BOOKS

1. Himmelblau, D. M. "Basic principles and calculations in Chemical Engineering", 8<sup>th</sup> Edition, PHI, 2013.
2. Foust, A. S. et al., "Principles of Unit Operations", 2<sup>nd</sup> Edition, John Wiley & Sons, 2014.
3. Coulson, J. M. and et al. "Coulson & Richardson's Chemical Engineering", 7<sup>th</sup> Edition, Vol. I & II, Butterworth-Heinemann (an imprint of Elsevier), 2011.
4. Robert W. Fox, Alan T. McDonald & Philip J. Pritchard "Introduction to Fluid Mechanics" 6<sup>th</sup> Edition, John Wiley & Sons 2003.

### EBOOKS/WEBLINKS

1. <http://www.pdfdrive.com/basic-principles-and-calculations-in-chemical-engineering-e185247644.html>
2. <http://www.pdfdrive.com/coulson-and-richardsons-chemical-engineering-fourth-edition-volume-3a-chemical-and-biochemical-reactors-and-reaction-engineering-e158316586.html>
3. <http://www.pdfdrive.com/stoichiometry-and-process-calculations-e187417539.html>

  
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Krishnagiri (DT), Tamilnadu.

**OBJECTIVES**

At the end of the course, the students should be able to:

- To understand about dimensional analysis and empirical methods governing the transport of momentum (fluid flow) in chemical and biotechnology engineering systems
- To analyze the scale-up of equipments for the production of biochemical products
- To assimilate the basic concepts of solid-liquid separation gained in earlier courses
- To predict various modes of heat transfer and exchange operations in transportation of fluids
- To understand the techniques of unit operations involved in designing a heat transfer in bioprocess equipment applications.

**UNIT I MIXING AND AGITATION**

9

Dimensional analysis-Rayleigh and Buckingham's method; principles of agitation, impellers, flow patterns: power consumption and power correlation in Newtonian liquids. Blending and mixing, agitator selection and scale up.

**UNIT II BASICS OF FILTRATION & CENTRIFUGATION**

9

Unit operations for solid-liquid separation - Filtration-Theory of filtration and equations; constant pressure, constant volume, constant rate filtration, discontinuous filter, continuous vacuum filter: rotary drum filters, Centrifugation-settling of solids, centrifuges, scale-up of centrifugation, centrifugal Filtration.

**UNIT III CONDUCTION HEAT TRANSFER**

9

Heat transfer phenomena-thermodynamics & heat transfer; Modes of heat transfer, Fourier's law of heat conduction, thermal conductivity; steady state conduction; Resistance concept- compound resistances in series, extended surfaces; unsteady state conduction; combined conduction and convection; 2dimensionalconduction.

**UNIT IV CONVECTION HEAT TRANSFER**

9

Convection-Forced and natural convection, Dimensional analysis, Dimensional numbers, Convection heat transfer coefficient, heat flux, individual heat transfer coefficients, overall heat transfer coefficients and fouling factors, application of dimensional analysis for convection, condensation phenomena, Film and dropwise condensation overtubes; heat transfer through boiling

**UNIT V HEAT EXCHANGERS**

9

Heat exchange equipment; counter current and parallel-current flows, LMTD correction factor, heat exchangers: single- pass 1-1 exchanger, 1-2 parallel-counterflow exchanger, 2-4 exchanger, multipass exchanger, enthalpy balances, and condensers-shell-tube condensers

**TOTAL HOURS 45 PERIODS**

**COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

- CO1: Understand about the transport of momentum (fluid flow) in chemical engineering systems  
 CO2: Improve their of knowledge in techniques of agitation, mixing of liquids, filtration operations and sedimentation separation  
 CO3: Understand modes of heat transferring techniques during extraction, distillation, evaporation  
 CO4: Evaluate effects of process variables while scaling up the bioprocess equipment and  
 CO5: Comprehend the important mechanical aspects while designing bioprocess equipment.

**TEXT BOOKS**

1. McCabe W.L., Smith J.C. Unit Operations in Chemical Engineering. 7<sup>th</sup> Edition. Mc graw hill 2014.


2. Dutta B.K, "Heat: Principles & applications", PHI publication 2000.
3. Gavahne.K.A., Unit Operations-I Fluid flow & mechanical separations, Niraliprakashan,2011.
4. Gavahne.K.A., Unit Operations-II Heat & Mass Transfer, Niraliprakashan, 25<sup>th</sup> edition, 2012.

#### REFERENCE BOOKS

1. Geankoplis C.J. Transport Processes and Unit Operations.4<sup>th</sup> edition, Prentice Hall India. 2003.
2. Coulson J.M., Richardson J.F., Backhurst J.R. and Harker J.M., "Coulson & Richardson's Chemical Engineering", 6<sup>th</sup> Edition, Vol.I & II, Butterworth-Heinman (an imprint of Elsevier), 2004.
3. Donald Q.Kern, "Process Heat Transfer", Tata Mc Graw Hill Book Co., New Delhi, 1997.
4. Foust, A.S. "Principles of Unit Operations", 2<sup>nd</sup> Edition, John Wiley & Sons, 1999.

#### EBOOKS/WEBLINKS

1. <https://www.pdfdrive.com/heat-and-mass-transfer-by-kothadaraman-e29924786.html>
2. <https://www.pdfdrive.com/fundamentals-of-heat-and-mass-transfer-6e-e14571835.html>
3. <https://www.pdfdrive.com/fluid-mechanics-heat-transfer-and-mass-transfer-chemical-engineering-practice-e157347975.html>
4. <https://www.pdfdrive.com/heat-and-mass-transfer-by-rk-rajput-e50661606.html>

  
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418BTT05 ENZYME TECHNOLOGY

L T P C

3 0 0 3

**Prerequisite** Biochemistry

## OBJECTIVES

At the end of the course, the students should be able to:

- To provide knowledge and application of working principles and their mechanism of action on enzymes
- To learn theoretical and practical aspects of kinetics
- To improve knowledge in the area of immobilization technique
- To learn enzyme reactions and their characteristics along with the production and purification process.
- To understand about the principles of Biosensors.

### UNIT I INTRODUCTION TO ENZYMES 9

*Nomenclature & Classification of enzymes. Mechanisms of enzyme action- Lock and key and induced fit model; concept to active site, catalysis, activator and inhibitors, specificity of enzyme action; Enzyme units; coenzymes, isoenzymes*

### UNIT II KINETICS OF ENZYMES 9

*Kinetics of single substrate reactions; Michelis–Menten equations, signification of Michelis – Menten equations, the lineweaver-burk plot, Eadie-hofstee and hanes plots: turn over number; types of inhibition–Competitive, uncompetitive and non-competitive inhibition; Allosteric regulation of enzymes; Monod, wymanmodel; pH and temperature effect on enzymes; Deactivation kinetics.*

### UNIT III ENZYME IMMOBILIZATION 9

*Physical and chemical techniques for enzyme immobilization–adsorption, matrix entrapment, encapsulation, cross-linking, covalent binding etc., - Examples, advantages and disadvantages of enzyme immobilization, Applications of immobilized enzyme systems.*

### UNIT IV PURIFICATION AND CHARACTERIZATION OF ENZYMES 9

*Production and purification of crude enzyme extracts from plant, animal and microbial sources; Molecular weight determination and characterization of enzymes; development of enzymatic assays.*

### UNIT V APPLICATION OF ENZYME BIOSENSORS IN INDUSTRY 9

*Enzyme biosensors; Definition and Main component of biosensor, Advantages and disadvantages of enzyme biosensors, Example of an Enzyme Biosensor-Electro chemical Biosensor, Blood Glucose Biosensor, Applications of biosensors in industry, Animal husbandry and health care and environment.*

**TOTAL HOURS 45 PERIODS**

## COURSE OUTCOMES

*Upon Completion of this course, students will be able to:*

- CO1: Develop knowledge on enzymes and enzyme reactions which is the key step towards understanding various concepts in biotechnology;
- CO2: Analyze theoretical and practical aspects of kinetics provide the importance towards interpreting the results;
- CO3: Apply the process for commercial production of enzymes;
- CO4: Implement ideas on processing, production and purification of enzymes on an industrial scale
- CO5: Design and novel biosensor products with better quality and wide commercial application.

## TEXT BOOKS


1. Harvey W. Blanch, Douglas S. Clark, "Biochemical Engineering", Marcel Dekker, Inc.2006
2. James M. Lee, "Biochemical Engineering", PHI, USA.2001
3. Nicholas C. Price and Lewis Stevens, "Fundamentals of Enzymology", Oxford university press 1999
4. Trevor Palmer "Enzymes: Biochemistry, Biotechnology and Clinical Chemistry" Horwood,

#### **REFERENCE BOOKS**

1. James. E. Bailey & David F. Ollis, "Biochemical Engineering Fundamentals", McGraw-Hill.2001
2. Wiseman, "Enzyme Biotechnology", Ellis Horwood Pub.2003
3. Faber K, Biotransformations in Organic Chemistry, IV edition, Springer
4. Roger Harrisonetal., "Bioseparation science and Engineering", Oxford University Press,2003.

#### **EBOOKS/WEBLINKS**

1. <https://nptel.ac.in/courses/102102033/>
2. <https://ndl.iitkgp.ac.in/>
3. <https://www.pdfdrive.com/microbial-enzyme-technology-in-food-applications-e185805089.html>
4. <https://www.pdfdrive.com/biosensors-and-biodetection-methods-and-protocols-volume-2-electrochemical-bioelectronic-piezoelectric-cellular-and-molecular-biosensors-e181167582.html>



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**Prerequisite** Environmental Science & Engineering

### OBJECTIVES

At the end of the course, the students should be able to:

- To understand the fundamentals of biotechnological concepts;
- To develop the skills in the area of environmental biotechnology and its pre-requisite(s) for PG studies in Biotechnology;
- To know the conversion of waste into energy using microorganisms;
- To understand about the eco-friendly bioproducts from renewable sources and
- To improve the skills in the area of waste water treatment technology.

#### UNIT I BIOGEOCHEMICAL ROLE OF SOIL MICROORGANISMS 9

Microbial flora of soil–Interactions among soil microorganisms–Nitrogen cycle–Carbon cycle–Sulfur cycle– Phosphorous cycle.

#### UNIT II BIODEGRADATION 9

Aerobic degradation of recalcitrant organic compounds by microorganisms– Growth associated degradation of aliphatic–Diversity of aromatic compounds–Co- metabolic degradation of organo pollutants – Degradative capacities of fungi. Anaerobic degradation of organic compounds – Degradation of hydrocarbons–Alkyl compounds–ketones–Aromatic compounds–Halogenated organics–Sulfonates–Nitro organics.

#### UNIT III BIOREMEDIATION TECHNOLOGIES 9

Remediation technologies–Bioventing–Biosparging and bioslurping–Phytoremediation–Bio Desulphurization of coal and oil–Microbial transformation of heavy metals–Bioleaching, bioaccumulation – Biosorption and bio precipitation of heavy metals.

#### UNIT IV ECO-FRIENDLY BIOPRODUCTS FROM RENEWABLE SOURCES 9

Fundamentals of composting process–Composting technologies–Composting systems–Compost quality–Biofertilizers–Biopesticides–Scientific aspects and prospects of biofuel production–Bioethanol–Bio hydrogen and biodiesel– *Biogas plant digester*.

#### UNIT V BIOLOGICAL TREATMENT OF WASTEWATER 9

Physical and chemical characteristics of waste water–Biological processes for waste water treatment- Activated sludge process–Trickling filter–Rotating biological contactors–Fluidized bed reactor– Upflow anaerobic sludge blanket reactor (UASB)–High-rate anaerobic waste water treatment.

**TOTAL HOURS 45 PERIODS**

### COURSE OUTCOMES

*Upon Completion of this course, students will be able to:*

- CO1: Develop and improve in standard of living;
- CO2: Understand the dynamic process integrated themes related to biodiversity;
- CO3: Envision the surrounding environment its function with technology;
- CO4: Understand the structure and biochemical aspects of various microbes and
- CO5: Acquire knowledge about the renewable forms of energy and its features of biomass and its utilization

### TEXT BOOKS

1. Jordening, H. J. and Winter, J., “Environmental Biotechnology: Concepts and Application”, Wiley- VCH Verlag, 2005.
2. Evans, G. M. and Furlong, J. C., “Environmental Biotechnology: Theory and Application”, John


- Wiley and Sons,  
3. 2003.  
4. Bhattacharya, B. C. and Banerjee, R., “Environmental Biotechnology”, Oxford University Press, 2007.  
5. Rajagopalan,R, ‘Environmental Studies-From Crisisto Cure’, Oxford University Press, 2005.  
6. G.Tyler Miller and Scott E. Spoolman,“Environmental Science”, Cengage Learning India PVT, Ltd., Delhi, 2014.

#### **REFERENCE BOOKS**

1. Pelczar, M.J., Chan, E. C. S. and Krieg, N.R., “Microbiology”, TataMcGraw-Hill,2005.
2. Rittmann, B. E. and McCarty, P. L., “Environmental Biotechnology: Principles and Applications”, McGraw-Hill, 2001.
3. Dharmendra S. Sengar, ‘Environmental law’, Prentice Hall of India Pvt., Ltd., New Delhi, 2007.
4. Erach Bharucha, “Text book of Environmental Studies”, Universities Press (I) Pvt, Ltd, Hyderabad,2015.

#### **EBOOKS/WEBLINKS**

1. <http://www.pdfdrive.com/environmental-biotechnology-principles-and-applications-e157042082.html>
2. <http://www.pdfdrive.com/environmental-science-e12033451.html>
3. <http://www.pdfdrive.com/environmental-biotechnology-theory-and-application-e7353867.html>

  
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**OBJECTIVES**

At the end of the course, the students should be able to:

- Gain knowledge on the basis of measurements and instruments used in Molecular Biology;
- Provide hands-on experience in performing basic molecularbiology techniques;
- Understand the theory behind each technique and to describe common applications of each methodology in biological research;
- Gain knowledge about separation of biological products;
- Take up specialized projects in Molecular biology which is a pre-requisite for research work.

**LIST OF EXPERIMENTS**

1. Preparation of reagents, handling equipments and lab safety in molecular biology labs;
2. Quantification of DNA using UV spectrophotometer;
3. Estimation of melting point (tm) of DNA;
4. Determination of molecular weight of DNA by Agarose gel electrophoresis;
5. Determination of protein profile by SDSPAGE;
6. Isolation of genomic DNA-Plant Cells;
7. Isolation of genomic DNA-Yeast Cells;
8. Isolation of DNA from whole blood;
9. Isolation of bacterial plasmid DNA;
10. Restriction enzyme digestion.

**TOTAL HOURS 45 PERIODS**

**COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

CO1: Demonstrate knowledge and understanding of the principles under pinning important techniques in molecular biology;

CO 2: Present advanced knowledge in the specialized fields of Molecular Biology;

CO 3: Demonstrate knowledge and understanding of applications of these techniques;

CO 4: Demonstrate the ability to carry out laboratory experiments and interpret the results;

CO5: Understand and be aware of hazardous chemicals and safety precautions in case of an emergency.

**TEXT BOOKS**

1. Sambrook, Joseph and David W. Russell "The Condensed Protocols: From Molecular Cloning: A Laboratory Manual" Cold Spring Harbor,2006.
2. David Freifelder,"MolecularBiology",4<sup>th</sup>revised Jones & Bartlett Publisher.2005.
3. Dr.P.K.Gupta, "Molecular Biology and Genetic Engineering";2<sup>nd</sup> Reprint. Rastogi Publications, 2011.

**REFERENCE BOOKS**

1. Michael P. Weiner "Genetic Variation: A Laboratory Manual" *Rain Dance Technologies*,2007.
2. Robert Schleif "Genetics and Molecular Biology" 2<sup>nd</sup> Edition. The Johns Hopkins University Press.1993.
3. Carson, Susan, "MolecularBiologyTechniques"3<sup>rd</sup> Edition, Elsevier.2012.

**EBOOKS/WEBLINKS**

1. <https://www.kopykitab.com/Cell-And-Molecular-Biology-A-Lab-Manual-by-K-V-Chaitanya>
2. [https://www.researchgate.net/publication/226072152\\_Basic\\_Techniques\\_in\\_Molecular\\_Biology](https://www.researchgate.net/publication/226072152_Basic_Techniques_in_Molecular_Biology)
3. [http://genome.tugraz.at/MolecularBiology/WS11\\_Chapter09\\_.pdf](http://genome.tugraz.at/MolecularBiology/WS11_Chapter09_.pdf)

**OBJECTIVES**

At the end of the course, the students should be able to:

- To gain knowledge on the basis of measurements and instruments;
- To have a practical hands-on experience on absorption spectroscopic methods;
- To gain knowledge about separation of biological products;
- To acquire experience in the purification by performing chromatography and
- To validate and analyze using spectrometric and microscopic techniques.

**LIST OF EXPERIMENTS**

1. Ultra violet and visible spectrometry instrumentation
2. Determination of maximum wavelength of  $\text{KMnO}_4$
3. Determination of maximum wavelength for copper sulphate
4. Finding the maximum wavelength of  $\text{Fe}^{3+}$ (1,10 phenanthroline) using UV spectrometry
5. Absorption spectrum of plant pigments
6. UVspectra of nucleic acids
7. Estimation of  $\text{SO}_4^{2-}$  by Nephelometer
8. Estimation of  $\text{Al}^{3+}$  by fluorimetry
9. Estimation of trace elements by flame photometry
10. Separation and Identification of amino acids using paper chromatography
11. Separation and Identification of amino acids using TLC
12. Chromatography analysis using gel chromatography

**TOTAL HOURS 45 PERIODS**

**COURSE OUTCOMES**

*Upon Completion of this course, students will be able to:*

- CO1: Understand calibration of instruments;  
 CO2: Acquire knowledge on separation techniques used for biological products;  
 CO3: Understand and apply optical methods;  
 CO4: Acquire knowledge on different chromatographic methods for separation of biological product  
 CO5: Acquire knowledge of purification by chromatography.

**TEXT BOOKS**


1. Textbook of Qualitative Inorganic Analysis, AIVogal, ELBSedition1987.
2. A Biologist guide to principles and techniques of practical biochemistry keith Wilson, Kenneth H Gouicing 3<sup>rd</sup> edition ELBS Series.
3. Hobert H Willard D.L. Merrit J.R.J. A Dean instrumental methods Analysis, CBS Publishers Distributors1992.
4. Electrochemical Methods by Bard Faulkner 2nd Edition Wiley Publishers 2006.
5. Biophysical Chemistry by Upadhyay 4th Edition by Himalaya Publishing House2007.

**REFERENCE BOOKS**

1. Instrumental Methods of Analysis. D. Skoog, 2000 5th Edition College Publishers.
2. Instrumental Methods of Chemical Analysis Galen N. Ewing 5th Edition McGraw Hill International 2006.
3. Introduction to Instrumental Analysis by Robert D Braun, Pharma Book Syndicate 2005.
4. Instrumental Methods of Chemical Analysis by HKaurPPMPublishers1999.

**EBOOKS/WEBLINKS**

1. <http://www.pdfdrive.com/instrumental-analysis-by-skoog-holler-crouch-2007-brookscole-e79362564.html>
2. <http://www.pdfdrive.com/chemical-analysis-modern-instrumentation-methods-and-techniques-e19446473.html>
3. <http://www.pdfdrive.com/vogels-qualitative-inorganic-analysis-5th-ed-e46819938.html>

  
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**Prerequisite** Biochemistry

### OBJECTIVES

At the end of the course, the students should be able to:

- To study about various parameters affecting the natural properties of enzymes.
- To provide hands on experience in enzyme production and purification techniques.
- Provide hands-on experience in performing enzyme production and purification techniques.
- To understand the students on enzyme characterization and immobilization methods.
- Introduce students to the theory behind in each technique and to describe common applications of each methodology in biological research. This will facilitate the students to take up specialized project in enzyme production and purification will
- Be a pre-requisite for research work.

### LIST OF EXPERIMENTS

1. Production of microbial enzymes
2. Partial purification of enzymes
3. Partial digestion of protein using enzyme-amylase, invertase, papain, pepsin
4. Effect of pH on enzyme activity.
5. Effect of temperature on enzyme activity
6. Effect of substrate concentration on enzyme activity
7. Determination of stability of enzyme activity.
8. Quantitative analysis of enzyme-amylase, invertase, papain, pepsin
9. Estimation of  $V_{max}$  and  $K_m$ .
10. Assaying of alkaline phosphatase activity
11. Enzyme immobilization–Gel entrapment
12. Immobilization of yeast cells as biocatalyst for the production of ethanol from sugar.

**TOTAL HOURS 45 PERIODS**

### COURSE OUTCOMES

*Upon Completion of this course, students will be able to:*

CO1: Demonstrate the activity of enzyme with various factors

CO 2: Learnt the various process of enzyme immobilization

CO3: Awareness about various kinetic studies on enzymes

CO4: Demonstrate the ability to carry out laboratory experiments and interpret the results.

CO5: Explain about Enzyme kinetics and characterization and how to use them for practical applications

### TEXT BOOKS

1. Practical Enzymology, 2<sup>nd</sup> Edition, By Hans Biss wange, Wiley-VCH Verlag GmbH & Co. KGaA, 2012.
2. Practical Biochemistry for Colleges by E. J. Wood, 1<sup>st</sup> Edition, Elsevier, 1989.
3. Enzymes in Industry: Production and Applications: W. Gerhartz, VCH Publishers, New York, 1990


### REFERENCE BOOKS

1. Enzyme Technology by M.F. Chaplin and C. Bucke, Cambridge University Press, Cambridge, 1990.
2. Bailey and Ollis, "Biochemical Engineering Fundamentals", McGraw Hill (2<sup>nd</sup> Ed.), 1986.

3. Shuler and Kargi, "Bioprocess Engineering ", Prentice Hall, 1992.

#### **EBOOKS/WEBLINKS**

1. <https://www.pdfdrive.com/enzyme-kinetics-and-mechanism-part-d-developments-in-enzyme-dynamics-e157727403.html>
2. <https://www.pdfdrive.com/enzyme-technologies-for-pharmaceutical-and-biotechnological-applications-e184251789.html>
3. <https://www.pdfdrive.com/enzyme-engineering-methods-and-protocols-e164853179.html>
4. <https://www.pdfdrive.com/enzyme-studies-e164429917.html>

  
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**ADHIYAMAAN COLLEGE OF ENGINEERING (AUTONOMOUS), HOSUR-635 109**

**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015  
Semester V**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
515BTT01	IMMUNOLOGY	3	0	0	3	50	50	100

**Prerequisite** Microbiology

*At the end of the course, the students should be able to:*

- To discuss the structure, functions and integration of immune system.
- To mechanisms involved in immune system development and responsiveness
- To explain various techniques of monoclonal and engineered antibodies
- To understand of activation of system of a complement. Receptors. Negative immunoregulation.
- To explain the antigen-antibody interactions and how the immune system is protecting the body from foreign pathogens.

**Course Objectives**

**UNIT I Introduction 9**

Cells and tissues of immune system; hematopoiesis; innate and acquired immunity; types of immune responses; antigens: chemical and molecular nature; haptens; adjuvants. Immunization and vaccines, Immunotechniques

**UNIT II Humoral Response 12**

Development, maturation, activation and differentiation of B cells: Clonal purification theory; Structure and functions of antibodies: Genes and generation of diversity; Hybridoma technology for production of monoclonal antibodies- Antibody engineering, Kinetics of antibody response

**UNIT III Cellular Response 10**

Development, maturation, activation and differentiation of T cells; and CMI (Cell mediate immunity), TCR;

Clonal Anergy; Antigen presenting cells: Macrophage, langerhan's cells and B cells- Antigen processing and presentation; Classes of MHC; MHC/HLA genetic loci; HLA alleles and diseases

**UNIT IV Immunity to Infection and Hypersensitivity Reactions 7**

Immune response to infections: viruses, bacteria, fungi and parasites; Cytokines; Complement systems; Hypersensitivity and their types.

**UNIT V Immunology of Transplantation, Autoimmunity And Tumor 7**

Transplantation: types of graft; mechanism of graft rejection; HVG and GVH rejection; immunological strategies to prevent graft rejection; *Autoimmune diseases and their mechanism* Tumor immunity.

**Total Hours 45**

**Course Outcomes** Upon Completion of this course, students will be able to get :

*CO1: Awareness of immune system structure and function*




- CO2: Awareness of immunity to various pathogens*  
*CO3: Awareness of cellular and molecular aspects of lymphocyte activation, homeostasis differentiation, and memory.*  
*CO4: Awareness of molecular basis of complex, cellular processes involved in inflammation and immunity, in states of health and disease*  
*CO5: Awareness of tumour allergy and hypersensitivity reactions*

#### **Text Books**

- 1 Ashim K. Chakravarthy, "Immunology", Tata McGraw- Hill, 2010
- 2 Richard A Goldsby, Thomas J Kindt, Barbara A Osborne and Janis Kuby. "Immunology" 5<sup>th</sup> Edition, W.H. Freeman & Co., 2005
- 3 Benjamin E. and Leskowitz S. Immunology A short Course, Wiley Liss NY, 2010
- 4 William E. Paul "Fundamental Immunology", 7<sup>th</sup> edition, Library of congress cataloguing in publications, 2013
- 5 Danny Altmann "Immunology", 12<sup>th</sup> edition, British Society of Immunology, 2017.

#### **Reference Books**

- 1 Roitt I Male, Brostoff. "Immunology", Mosby Publ., 2017
- 2 Janeway, Travers, Walport and Shlomichik, (2001), "Immunobiology", Garland Publ., 2011
- 3 Ian R. Tizard. "Immunology- An Introduction" 4<sup>th</sup> Edition. Thomson Publ., 2013
- 4 Andrew H. Lichtman, Shiv Pilla, Abul K. Abbas, Cellular & Molecular Immunology, 7<sup>th</sup> edition, south Asia Publication, 2011
- 5 Dr.S.K.Gupta, "Essentials of Immunology", 2<sup>nd</sup> edition, Arya Publications, 2010.

  
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**ADHIYAMAAN COLLEGE OF ENGINEERING (AUTONOMOUS), HOSUR-635 109**

Department **BIOTECHNOLOGY** Programme **B. TECH. - BT** Regulation **2015**  
Semester **V**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
<b>515BTT02</b>	<b>GENETIC ENGINEERING</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>	<b>50</b>	<b>50</b>	<b>100</b>

**Prerequisite** **Molecular Biology**

*At the end of the course ,the students should be able to:*

- Course Objectives**
- *To understand the principle of nucleic acid isolation.*
  - *To understand the principles of PCR and their uses in genetic engineering.*
  - *To gain a thorough knowledge about nucleic acid hybridization.*
  - *To learn history of DNA sequencing and current methods and gene synthesis*
  - *To understand the genetic hypothesis*

**UNIT I      BASICS OF RECOMBINANT DNA TECHNOLOGY      9**

Introduction of recombinant DNA into host cells, manipulation of DNA – Restriction and modification enzymes, Design of linkers and adaptors; Characteristics of cloning and vectors; prokaryotic and eukaryotic host systems.

**UNIT II      POLYMERASE CHAIN REACTION TECHNIQUES      9**

Principle of polymerase chain reaction (PCR) - Components of PCR reaction and optimization of PCR - Gene specific primer and degenerate primer – Inverse PCR, Hot-start PCR, Loop mediated PCR -, Reversetranscription PCR and Real time PCR.

**UNIT III      PROTEIN TECHNIQUES      9**

Electrophoresis of protein – native and denaturing conditions, capillary and gel electrophoresis, 2D gel electrophoresis, Enzyme-linked immunosorbent assay, yeast hybrid system – one hybrid system – two hybrid system, phage display.

**UNIT IV      TESTING OF GENETIC HYPOTHESIS      9**

Probability in the prediction of progeny distributions using Binomial distribution, Genetic hypothesis, Chi-square method, Genetic analysis of quantitative traits using Statistics (Mean, Variance, Standard deviation, Correlation, Regression), Heritability is useful in predicting the phenotypes of offspring.

**UNIT V      TRANSGENIC TECHNOLOGY      9**

Principles of Transgene Technology. Scope of Transgenetic Technology. Gene tagging (T-DNA tagging and Transposon tagging) in gene analysis (identification and isolation of gene), Transgenic and Gene Knockouts Technologies-Targeted gene replacement, Chromosome engineering.

**Total Hours      45**

Upon Completion of this course, students will be able to :

**Course  
Outcomes**

CO:1. Understand the basics of biotechnology

CO:2 Understand the value of and the processes involved with the polymerase chain reaction (PCR).

CO:3 Understand the concept of recombinant DNA technology or genetic engineering

CO:4 Analyze a research problem and step-by-step instructions for conducting experiments or testing hypothesis

CO:5 Explain the general principles of generating transgenic plants, animals and Genetically modified organisms.

**Text Books**

1

Klug, Cummings and Spencer. "Concepts of Genetics" published Pearson, 2016.

2

Daniel L. Hartl, Maryellen Ruvolo. "Genetics: Analysis of Genes and Genomes" 8<sup>th</sup> Edition, Published Laxmi (Pvt. Ltd). 2011.

**Reference Books**

1

Gardner, Simmons and Snustad. "Principles of Genetics" 8<sup>th</sup> Edition, Published, Wiley. 2006

2

Benjamin A. Pierce. "Genetics: A Conceptual Approach" 4<sup>th</sup> Edition, Published, W H Freeman & Co. 2010.

3

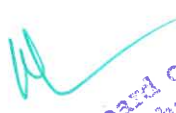
Scott F. Gilbert and Susan R. Singer. "Developmental Biology (Developmental Biology Developmental Biology)" 9<sup>th</sup> Edition, Published, Sinauer Associates, 2010.

4

Robert J. Brooke, "Genetics: Analysis and Principles" 4<sup>th</sup> Edition, McGraw-Hill Higher Education, 2012.

5

Smita Rastogi and Neelam Pathak. "Genetic Engineering (Oxford Higher Education)" 1<sup>st</sup> Edition, Oxford University Press, 2009.

  
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**ADHIYAMAAN COLLEGE OF ENGINEERING (AUTONOMOUS), HOSUR-635 109**

**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015**  
**Semester V**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
<b>515BTT03</b>	<b>BIOPROCESS ENGINEERING I</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>	<b>50</b>	<b>50</b>	<b>100</b>

**Prerequisite** Microbiology, Basic Industrial Biotechnology

*At the end of the course, the students should be able to:*

**Course Objectives**

- *To study the historical development of bio process technology, design of fermenter and types of fermentation process*
- *To gain knowledge about formulation, optimization of medium and principles of sterilization*
- *To inculcate the stoichiometry and energetics of cell growth and product formation*
- *To evaluate the kinetics and mechanism of microbial growth*
- *To gain knowledge related to the processing of fermentor*

**UNIT I OVERVIEW OF FERMENTATION PROCESSES 9**

Introduction to bioprocessing: Historical development of Bioprocess technologies, General requirements of fermentation processes, Basic design and construction of fermenters and ancillaries, Main parameters to be monitored and controlled in fermentation processes. Solid-state fermentation and its applications.

**UNIT II RAW MATERIALS AND MEDIA DESIGN FOR FERMENTATION PROCESS 9**

Criteria for good medium, medium requirements for fermentation processes, carbon, nitrogen, minerals, vitamins and other complex nutrients, oxygen requirements, Medium Formulation: Types of media – media design and usage of various commercial media for industrial fermentations, Medium optimization.

**UNIT III STERILIZATION KINETICS 8**

Thermal death kinetics of microorganisms, Batch and continuous heat sterilization of liquid media, Filter sterilization of liquid media, Air sterilization and design of sterilization equipment.

**UNIT IV METABOLIC STOICHIOMETRY AND ENERGETICS 10**

Stoichiometry of cell growth and product formation: Elemental balances, degrees of reduction of substrate and biomass, available electron balances, yield coefficients of biomass and product formation, maintenance coefficients. Energetic analysis of microbial growth and product formation: Oxygen consumption and heat evolution in aerobic cultures, Thermodynamic efficiency of growth.

**UNIT V KINETICS OF MICROBIAL GROWTH AND PRODUCT FORMATION 10**

Phases of Cell growth in batch cultures, Simple unstructured kinetic models for microbial growth, growth of filamentous organisms, product formation kinetics – Leudeking - Piret models, substrate and product inhibition on cell growth and product formation.

**Total Hours 45**

*Upon Completion of this course, students will be able to :*

**Course**

**Outcomes**

*CO1: Develop skills of the students in the area of bioprocess technology with the emphasis on bioprocess principles*

*CO 2: Discuss and distinguish the medium requirements and optimization methods*

*CO 3: Explain the sterilization kinetics of medium and equipments*

*CO 4: Learn about fermentation processes, metabolic stoichiometry, energetics, kinetics of microbial growth etc*


*CO 5: Understand the kinetics of microbial growth that plays a vital role in the fermentation process*

#### **Text Books**

- 1 Pauline. M. Doran, "Bioprocess Engineering Principles", Academic press, 2012.
- 2 Stanbury. P. F, Whitaker. A and Hall. S. J, "Principles of Fermentation Technology", 2<sup>nd</sup> Edition, Butterworth– Heinemann, 1995.

#### **Reference Books**

- 1 Najafpour. G. D, "Biochemical Engineering and Biotechnology", Elsevier, 2007.
- 2 Shuler. M. L and Kargi. F, "Bioprocess Engineering: Basic Concepts" 2<sup>nd</sup> Edition, Pearson, 2002.
- 3 Bailey. J. E and Ollis. D. F, "Biochemical Engineering Fundamentals", 2<sup>nd</sup> Edition, McGraw-Hill, 2010.
- 4 Blanch. H. W and Clark. D. S, "Biochemical Engineering". Marcel & Dekker, Inc., 2007.
- 5 Rao. D. G, "Introduction to Biochemical engineering" ", 2<sup>nd</sup> Edition, McGraw-Hill, 2010.



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**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015**  
**Semester V**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
<b>515BTT04</b>	<b>Fundamentals of Mass Transfer</b>	<b>3</b>	<b>1</b>	<b>0</b>	<b>4</b>	<b>50</b>	<b>50</b>	<b>100</b>

**Prerequisite** Fundamentals of Unit Operations

*At the end of the course, the students should be able to:*

- *Explain the basic principles of mass transfer operations and other separation processes with examples.*
- *Impart knowledge on how certain substances undergo the physical change with diffusion/mass transfer of components from one phase to other phases.*
- *Focus on absorption and distillation operations and the process design aspects of the same operations.*
- *Understand extraction and leaching operations and their applications in bioprocessing industry.*
- *Understand adsorption and drying operations and the process design aspects of the same operations.*

**Course Objectives**

**UNIT I DIFFUSION AND MASS TRANSFER 9+3**

Diffusion: Molecular diffusion, Fick's law of diffusion, steady state molecular diffusion in gases and liquid one component transferring to non diffusing component and equimolar diffusivity estimation, Inter phase Mass Transfer; Mass Transfer coefficients, Concept of overall mass transfer coefficient for liquids and gases, diffusivity measurement and prediction.

**UNIT II GAS LIQUID OPERATIONS 9+3**

Principles of gas absorption; Single component absorption in single and multistage operation; selection criteria for solvents, material balance, minimum gas-liquid ratio, Design principles of packed absorbers- HETP, HTU and NTU concepts, Industrial absorbers.

**UNIT III VAPOUR LIQUID OPERATIONS 9+3**

V-L Equilibria, P-x-y and T-x-y diagrams, relative volatility, Raoult's law; Ideal behavior of fluids, types of Distillation-Simple, Steam and Flash Distillation; Continuous distillation; Design calculations- McCabe-Thiele method, Concept of minimum, total and optimum reflux ratio, deviations from ideality - Extractive distillation and Azeotropic distillation.

**UNIT IV EXTRACTION OPERATIONS 9+3**

Liquid- liquid extraction: distribution coefficient, ternary systems and triangular diagrams, solvent selection criteria for extraction, single stage and multistage extraction – immiscible system, extraction equipments. Solid- liquid equilibria, Leaching Principles, constant underflow staged processes - Single stage leaching, multistage counter current leaching, Leaching equipments – Batch and continuous types.

**UNIT V SOLID FLUID OPERATIONS 9+3**

Types of adsorption, Nature of adsorbents, Langmuir and Freundlich isotherm, calculation of staged processes, adsorption equipments – Batch and fixed bed adsorption; Drying - Mechanism, Drying curves-Time of Drying calculation; Batch and continuous drying equipments.

**Total Hours 60**

*Upon Completion of this course, students will be able to :*

**Course Outcomes**

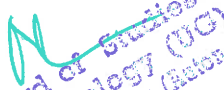
- CO1: define the basic principles of mass transfer operations and the measurement of diffusivity, mass transfer coefficient;*
- CO2: understand the importance of mass transfer phenomena in the design of process equipment in distillation operations;*
- CO 3: understand the HETP, NTU and HTU concepts of various gas absorption packed tower columns;*
- CO 4: understand the design aspects of extraction and various leaching equipments and*
- CO 5: understand the importance of adsorption and drying processes and their industrial applications.*

**Text Books**

- 1 Geankoplis C J. “Transport process and separation process principles”, 4<sup>th</sup> edition, Prentice Hall of India. 2003
- 2 Anantharaman N. and Meera Sheriffa Begum K.M. “Mass Transfer - Theory and Practice”, New Delhi: PHI Learning Private Limited. 2011
- 3 Treybal R.E. Mass Transfer Operations. 3<sup>rd</sup> edition. McGraw-Hill, 1981.

**Reference Books**

- 1 Warren L. McCabe, Julian C. Smith, Peter Harriot. “Unit Operations of Chemical Engineering”, 7<sup>th</sup> edition, New Delhi: McGraw Hill. 2012
- 2 Ghosal, S. K., Sanyal S. K. & Datta S.. “Introduction to Chemical Engineering”, New Delhi: Tata McGraw Hill. 2006
- 3 Benitez J, Principles and modern applications of Mass Transfer Operation, Wiley, 2009.
- 4 Coulson and Richardson, “Chemical Engineering”. Vol I & II, New Delhi: Asian Books Pvt Ltd, 1998.

  
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**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015**  
**Semester V**

515BTT05	CHEMICAL THERMODYNAMICS AND BIOTHERMODYNAMICS	Hours/Week			Credit C	Maximum Marks		
		L	T	P		CA	EA	Total
		3	0	0	3	50	50	100

**Prerequisite** Stoichiometry and process calculations

*At the end of the course the students should able to*

**Course Objectives**

- To study about the ideal and non-ideal behavior properties of fluids
- To understand about the determination of solution on thermodynamic properties
- To deal thermodynamic properties of fluids on its equilibrium in phase change
- To deal thermodynamic properties of fluids on its chemical reaction under equilibrium condition
- To analyse the energy in process on behavior with its properties

**UNIT I THERMODYNAMIC PROPERTIES OF FLUIDS 9**

Basics concepts in thermodynamics, Volumetric properties of fluids exhibiting non ideal behaviour; residual properties; estimation of thermodynamic properties using equations of state; calculations involving actual property exchanges; Gibbs Helmholtz Equation, Maxwell's relations and applications.

**UNIT II SOLUTION THERMODYNAMICS 9**

Partial molar properties; concepts of chemical potential and fugacity; ideal and non-ideal solutions; concepts and applications of excess properties of mixtures; activity coefficient; composition models; Gibbs Duhem equation.

**UNIT III PHASE EQUILIBRIA 9**

Criteria for phase equilibria; v-l-e calculations for binary and multi component systems; Bubble point, Dew point Calculation, liquid-liquid equilibria and solid-solid equilibria.

**UNIT IV CHEMICAL REACTION EQUILIBRIA 9**

Equilibrium criteria for homogeneous chemical reactions; evaluation of equilibrium constant; effect of temperature and pressure on equilibrium constant; calculation of equilibrium conversion and yields for single and multiple reactions.

**UNIT V THERMODYNAMIC ANALYSIS OF PROCESSES 9**

Concept of lost work; entropy generation; calculation of real irreversible processes; power cycle; liquefaction, Carnot Cycle, Biothermodynamics.

**Total Hours 45**

*Upon completion of this course, students will be able to*

**Course Outcomes**

- CO:1 Knowledge on ideal and non-ideal behaviour in thermodynamics on properties of fluids
- CO:2 Knowledge on solutions thermodynamics to determine the properties in the processes.
- CO:3 Description of properties criteria in order to maintain the phase change coexist in equilibrium
- CO:4 Description of properties criteria in order to maintain the chemical reactions coexist in equilibrium




*CO: 5 Knowledge on energy utilization and to interpret thermodynamic properties data in the bio processing operations.*

**Text Books**

- 1 Narayanan K.V. A Text Book Of Chemical Engineering Thermodynamics. Prentice Hall India, Eighth Edition 2013.
- 2 Smith J.M., Van Ness H.C., Abbot M.M. Chemical Engineering Thermodynamics. 6<sup>th</sup> Edition. McGraw-Hill, 2005

**Reference Books**

- 1 Sandler S.I. Chemical and Engineering Thermodynamics. John Wiley, 3 edition 1998.
- 2 B.G.Kyle, "Chemical process thermodynamics", 2<sup>nd</sup> Edn., Prentice Hall of India Pvt.Ltd., New Delhi 2000.

  
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**Department** BIOTECHNOLOGY      **Programme** B. TECH. - BT      **Regulation** 2015  
**Semester V**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
515BTP07	IMMUNOLOGY LABORATORY	0	0	4	2	50	50	100

**Prerequisite** Microbiology, cell biology lab

*At the end of the course, the students should be able to:*

**Course Objectives**

- To state the principle of the routine immunologic procedures performed in the clinical laboratory.
- To describe the immunologic responses involved in preventing and combating infections.
- To understand the role of antibodies in the immune response
- To understand the molecular specificity of antibodies for specific antigens
- To simulate the spread of an infectious disease and determine its source with an ELISA assay

**LIST OF EXPERIMENTS**

1. Handling of animals, immunization and raising antisera
2. Identification of Blood cells
3. Differential count of white blood cells
4. Blood grouping (ABO & Rh factor)
5. Widal Test (Slide & Tube Test)
6. Ouchterlony double immune diffusion technique (ODD)
7. Radial immune diffusion (RID) (*mancini method*)
8. Immunoelectrophoresis
9. Isolation of monocytes from blood
10. Isolation of peripheral blood mononuclear cells Identification of T cells by T cell resetting using sheep RBC.
11. Enzyme Linked Immuno Sorbent Assay
12. Western Blotting

**Total Hours 45**

*Upon Completion of this course, students will be able to get :*

**Course Outcomes**

*CO 1: Awareness of basic and state-of-the-art experimental methods and technologies CO2: Awareness to develop an ability to summarize, integrate and organize information and relate it to disease outcomes*

*CO3: Awareness to evaluate the potential for current research and new discoveries to improve our understanding of immunology and its relevance to human health and to our society.*

*CO4: Awareness to use medical case reports, identify “disease defects” and define molecular or cellular targets for therapeutic intervention:*


*CO5: Awareness to understand basic mechanisms and preventive therapeutic implications*

#### **Text Books**

1. Ashim K. Chakravarty, "Immunology", Tata McGraw- Hill, 2010
2. Richard A Goldsby, Thomas J Kindt, Barbara A Osborne and Janis Kuby. "Immunology" 5<sup>th</sup> Edition, W.H. Freeman & Co., 2005
3. Benjamin E. and Leskowitz S. "Immunology A short Course", Wiley Liss NY, 2010
4. Mark Peakman and Leonie Taams, "Clinical & Experimental Immunology", 12<sup>th</sup> edition, British Society for Immunology, 2017.
5. Frank C. Hay, Olwyn M. R. Westwood "Practical Immunology", 4<sup>th</sup> Edition Wiley Blackwell Publications, 2010

#### **Reference Books**

- 1 Talwar, G.P and Gupta, S.K. 2004" A Handbook of practical and immunology", CBS Publishers & Distributors.
- 2 Janeway, Travers, Walport and Shlomichik, (2001), "Immunobiology", Garland Publ., 2011
- 3 Ian R. Tizard. "Immunology- An Introduction. 4<sup>th</sup> Edition". Thomson Publ., 2013
- 4 J Ochei and A. Kolhatkar "Medical Laboratory Science Theory and Practice" by PPM Publishers. 1999.
- 5 Barbara Detrick, Robert G. Hamilton, John L. Schmitz "Manual of Molecular and Clinical Laboratory Immunology", 8<sup>th</sup> edition ASM Press, 2016

  
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**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015**  
**Semester V**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		CA	EA	Total
515BTP08	GENETIC ENGINEERING	0	0	4	2	50	50	100

**LABORATORY**

**Prerequisite** Molecular biology lab

*At the end of the course ,the students should be able to:*

**Course Objectives**

- *To illustrate creative use of modern tools and techniques for manipulation and analysis of genomic sequences.*
- *To expose students to application of recombinant DNA technology in biotechnological research.*
- *To understand research methodologies employing genetic engineering techniques.*
- *To understand the principles of PCR and their uses in genetic engineering.*
- *To understand the principles of blotting*

**LIST OF EXPERIMENTS**

1. Isolation of plasmid DNA
2. Restriction enzyme digestion
3. Purification of digested DNA- Gel Elution
4. Preparation of competent cells
5. Transformation and screening in *E. coli*
6.  $\beta$ -galactosidase assay
7. DNA cloning
8. PCR
9. DNA finger printing
10. SDS-PAGE
11. Western blotting
12. Southern blotting

**Total Hours**

**45**

*Upon Completion of this course, students will be able to :*

**Course Outcomes**

- CO1: Technical know-how on versatile techniques in recombinant DNA technology. CO2: An ability to design and conduct experiments, as well as to analyze and interpret data
- CO3: Apply of genetic engineering techniques in basic and applied experimental biology.
- CO4: Develop proficiency in designing and conducting experiments involving genetic manipulation.
- CO5: An ability to use the techniques, skills, and modern engineering tools necessary for engineering practice.

**Text Books**

Isil Aksan Kurnaz, “Techniques in Genetic Engineering” Published, CRC Press, 2015

David Micklos "Genome science a practical and conceptual introduction to molecular genetic analysis in eukaryotes" 1<sup>st</sup> Edition, Published, Cold Spring. 2013


Rolf H. J. Schlegel, "Rye: Genetics, Breeding, and Cultivation" Published, CRC Press. 2013

T A Brown "Introduction to Genetics: A Molecular Approach" Published, Garland Science. 2011.

*Setlow, Jane K. "Genetic Engineering- Principles and Methods" Published, Plenum. 2003*

#### Reference Books

- 1 Isil Aksan Kurnaz, "Techniques in Genetic Engineering" Published, CRC Press. 2015.
- 2 DR. P.S. VERMA and V K Agarwal. "Genetic Engineering" Published, S. Chand Publishing. 2009.
- 3 Utpal Roy and Vishal Saxena. "A Handbook of Genetic Engineering" 47<sup>th</sup>, Edition, Published, Kalyani. 2007.
- 4 Vennisonand S John. "Laboratory Manual for Genetic Engineering" published, Prentice Hall India Learning Private Limited. 2009.
- 5 C.C. Girianand Archana Giri. "Plant Biotechnology: Practical Manual" Published, I K International Publishing House Pvt. Ltd. 2007

  
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**Department** BIOTECHNOLOGY      **Programme** B. TECH. - BT      **Regulation** 2015  
**Semester V**

515BTP09	BIOPROCESS ENGINEERING LABORATORY - I	Hours/Week			Credit	Maximum Marks		
		L	T	P	C	CA	EA	Total
		0	0	4	2	50	50	100

**Prerequisite** Bioprocess principles theory

*At the end of the course the students should able*

**Course Objectives**

- To develop basic experimental skills for preparing medium and sterilization.
- To provide practical understanding of effect on parameters on cell growth
- To provide training on upstream processes technology
- To provide knowledge on preparation and utilization offer mentor
- To provide knowledge on production primary and secondary metabolite.

**LIST OF EXPERIMENTS**

1. Preparation of bioreactor, utilizes for bioreactor
2. Medium preparation and sterilization
3. Effect of temperature on cell growth
4. Effect of pH on cell growth
5. Monod kinetics
6. Growth of bacteria-Estimation of biomass, calculations of specific growth rate, yield coefficient
7. Growth of Yeast-Estimation of biomass, calculations of specific growth rate, yield coefficient
8. Effect of substrate inhibition on cell growth
9. Production of primary metabolites
10. Production of secondary metabolites
11. Medium optimization-Plackett burmandesign
12. Medium optimization-Response surface methodology
13. Single cell protein (SCP) production by continuous culture

**TOTAL HOURS 45**

*Upon completion of this course, students will be able to have*

**Course Outcomes**

- CO:1 Knowledge on preparation of medium and sterilization in upstream processes*  
*CO:2 Knowledge on optimization of cell growth*  
*CO:3 Exposure to upstream processes and preparation before the fermentation*  
*CO:4 Knowledge on preparation and utility of bioreactor*  
*CO:5 Knowledge on production of metabolites in lab scale fermentor*

**Text Books**

- 1 S. Kulandaivelu and S. Janarthanan,"Practical Manual on Fermentation Technology" IK International publishing house, NewDelhi ,2012
- 2 Palvannan T, Shanmugam S, Satish Kumar T,"Laboratory Manual On Biochemistry, Bioprocess & Microbiology", Scitech Publications (India) Pvt Lt ,2006

### Reference Books

- 1 Sarfaraz K. Niazi, Justin L. Brown, "Fundamentals of Modern Bioprocessing" CRC Press, 2015
- 2 Pauline Doran, "Bioprocess Engineering Calculation", Blackwell Scientific Publications, 1998.

  
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**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015  
Semester VI**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		CA	EA	Total
<b>615BTT01</b>	<b>BIOINFORMATICS</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>	<b>50</b>	<b>50</b>	<b>100</b>

**Prerequisite** Basics of computing and C programming

*At the end of the course, the students should be able to:*

- Course Objectives**
- To develop interdisciplinary skills in the applications of computers in biotechnology.
  - To navigate through internet-based biological databases and genomic browsers
  - To let the students know the recent evolution in biological science.
  - To develop the knowledge related to phylogenetic tree
  - To gain knowledge related to tools used in bioinformatics

**UNIT I INTRODUCTION 9**

Introduction to Bioinformatics – applications, Operating systems- types, Elementary UNIX commands, TCP/IP, Telnet, FTP, Protocols, Hardwares, Network topology, Search engines.

**UNIT II BIOLOGICAL DATABASES 9**

Introduction to databases – Data life cycle Biological databases; Primary nucleotide databases (EMBL, GeneBank and DDBJ); Primary protein databases (SwissProt) Secondary protein databases (PROSITE); Structural databases – SCOP and CATH. Sequence retrieval from database

**UNIT III PATTERN MATCHING AND DYNAMIC PROGRAMMING 9**

Introduction to pairwise sequence alignment – local vs. global; Dynamic programming – Needleman – Wunsch algorithm & Smith – Waterman algorithm; Dot matrix analysis; substitution matrices, BLAST – FASTA – Statistical methods – Hidden Markov models.

**UNIT IV PHYLOGENY 9**

Introduction to multiple sequence alignment, Introduction; mutations; mutations as a measure of time; Phylogenetic analysis Distance matrix methods, character based methods. Molecular clock theory, Bootstrapping.

**UNIT V ADVANCED TOPICS IN BIOINFORMATICS 9**

Introduction to Systems Biology and Synthetic Biology, Microarray analysis - types and applications, Bioinformatics approaches for drug discovery.

**Total Hours 45**

*Upon Completion of this course, students will be able to get :*

- Course Outcomes**
- CO1: Develop bioinformatics tools with programming skills.  
 CO2: Apply computational based solutions for biological perspectives  
 CO3: Pursue higher education in this field.  
 CO4: Practice life-long learning of applied biological science.  
 CO5: Developed the knowledge related to phylogenetic tree



### **Text Books**

- 1 Lesk, A. K., "Introduction to Bioinformatics" 4<sup>th</sup> Edition, Oxford University Press, 2013
- 2 Dan Gusfield, "Algorithms on Strings, Trees and Sequences: Computer Science and Computational Biology" Cambridge University Press, 1997.
- 3 Durbin, R., Eddy, S., Krogh, A., and Mitchison, G., "Biological Sequence Analysis Probabilistic Models of proteins and nucleic acids" Cambridge, UK: Cambridge University Press, 1998.
- 4 Mount, D.W., "Bioinformatics Sequence and Genome Analysis" 2<sup>nd</sup> Edition, Cold Spring Harbor Laboratory Press, 2004
- 5 Bergeron, B. Bioinformatics Computing, 2<sup>nd</sup> Edition, Prentice Hall of India Learning Pvt (Ltd), India, (2009).

### **Reference Books**

- 1 Attwood, T.K and Parry Smith, D.J. Introduction to Bioinformatics, 1<sup>st</sup> Edition, Pearson Education Asia, India, (2002).
- 2 Uri Alon, An Introduction to Systems Biology: Design Principles of Biological Circuits, Chapman & Hall, 2006.
- 3 Andreas D. Baxevanis, B. F. Francis Ouellette: Bioinformatics: A Practical Guide to the Analysis of Genes and Proteins, Volume 39, John Wiley, 1998
- 4 Baldi, P. and Brunak, S., "Bioinformatics: The Machine Learning Approach" 2<sup>nd</sup> Edition, MIT Press, 2001.
- 5 J. Pevsner, Bioinformatics and Functional Genomics, 2<sup>nd</sup> Edn., Wiley-Blackwell, 2009.

  
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**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015**  
**Semester VI**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		CA	EA	Total
615BTT02	CHEMICAL REACTION ENGINEERING	3	1	0	4	50	50	100

**Prerequisite** Stoichiometric and process calculations

*At the end of the course, the students should be able to:*

**Course Objectives**

- To impart the basic concepts in reaction kinetics
- To provide the information about different reactor systems and deriving the performance equations for different reactor systems.
- To develop knowledge for design of ideal reactors and RTD studies
- To acquire knowledge in heterogeneous reactions and reactor types.
- To gain knowledge related to various types of reactor

**UNIT I SCOPE OF CHEMICAL KINETICS & CHEMICAL REACTION ENGINEERING**

**9+3**

Introduction to Chemical kinetics; rate equation, rate constant, elementary and non- elementary reactions; concentration and temperature dependence; development of rate equations for different homogeneous reactions,

Search for reaction mechanism; Interpretation of batch reactor data-Integral and differential method of analysis (constant volume batch reactor).

**UNIT II IDEAL REACTORS**

**9+3**

Isothermal batch, flow, semi-batch reactors; performance equations for single reactors - batch, plug flow and mixed flow reactors; space time and space velocity; multiple reactor systems; multiple reactions.

**UNIT III IDEAL FLOW AND NON IDEAL FLOW**

**9+3**

Basics of non ideal flow; RTD function and measurement, RTD in plug flow and mixed flow reactor, relation among E, F and C curve, conversion in non ideal flow, non-ideal flow models- tank in series and dispersion models; reactor performance with non-ideal flow.

**UNIT IV UNIT IV GAS-SOLID, GAS-LIQUID REACTIONS**

**9+3**

Resistances and rate equations; heterogeneous catalysis; reactions steps; selection of a model, unreacted core models for spherical particles - progressive conversion model and shrinking core model, determination of rate controlling step.

**UNIT V****FIXED BED AND FLUID BED REACTORS**

9+3

G/L reactions on solid catalysis; trickle bed, slurry reactors; three phase-fluidized beds; reactors for fluid-fluid reactions; tank reactors.

**Total Hours 60**

**Course Outcomes**

*Upon Completion of this course, students will be able to get :*

*CO1: Write the rate equation for any type of chemical reaction.*

*CO 2: Relate and calculate the conversions, concentrations and rates in a reaction and identify, formulate and solve chemical engineering problems.*

*CO 3: Design reactors for heterogeneous reactions and optimize operating conditions.*

*CO 4 understood the concept of RTD*

*CO 5 gained knowledge about the reaction catalysis*

**Text Books**

- 1 Levenspiel O. Chemical Reaction Engineering. 3<sup>rd</sup> Edition. John Wiley.1999.
- 2 Fogler H.S. Elements of Chemical Reaction Engineering. Prentice Hall India.2002
- 3 Mark E.Davis and Robert J.Davis, Fundamentals of Chemical Reaction Engineering, McGraw-Hill Higher Education; 1<sup>st</sup> edition 2002

**Reference Books**

- 1 Missen R. W., Mims C. A., Saville B. A. Introduction to Chemical Reaction Engineering and Kinetics. John Wiley.1999
- 2 Dawande, S.D., "Principles of Reaction Engineering", 1<sup>st</sup> Edition, Central Techno Publications, 2001.
- 3 Richardson, J.F. and Peacock, D.G., "Coulson Richardson - Chemical Engineering", Vol.III, III<sup>rd</sup> Edition, Butterworth- Heinemann- Elsevier, 2006.

  
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**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015**  
**Semester VI**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		CA	EA	Total
<b>615BTT03</b>	<b>BIOPROCESS ENGINEERING-II</b>	<b>3</b>	<b>1</b>	<b>0</b>	<b>4</b>	<b>50</b>	<b>50</b>	<b>100</b>

**Prerequisite** Bioprocess Engineering – I and Fundamentals of Mass Transfer

*At the end of the course, the students should be able to:*

**Course Objectives**

- *To impart the basic of different operational modes of bioreactors*
- *To develop knowledge for design aspects of bioreactors scale up for various systems*
- *To acquire knowledge in reactor consideration for enzyme systems and modeling and simulation of bioprocess.*
- *To develop knowledge in recombinant cultivation systems.*
- *To gain knowledge about the scale up process*

**UNIT I OPERATIONAL MODES OF BIOREACTORS 9+3**

Fed batch cultivation, Cell recycle cultivation, Cell recycle cultivation in waste water treatment, twostage cultivation Packed bed reactor, airlift reactor, fluidized bed reactor bubble column reactors.

**UNIT II BIOREACTOR SCALE – UP 9+3**

Regime analysis of bioreactor processes, oxygen mass transfer in bioreactors - microbial oxygen demands; methods for the determination of mass transfer coefficients; mass transfer correlations. Scale up criteria for bioreactors based on oxygen transfer, power consumption and impeller tip speed.

**UNIT III BIOREACTOR CONSIDERATION IN ENZYME SYSTEMS 9+3**

Analysis of film and pore diffusion effects on kinetics of immobilized enzyme reactions; formulation of dimensionless groups and calculation of effectiveness factors. Design of immobilized enzyme reactors – packed bed, fluidized bed and membrane reactors.

**UNIT IV MODELLING AND SIMULATION OF BIOPROCESSES 9+3**

Study of structured models for analysis of various bioprocess – compartmental models, models of cellular energetics and metabolism, single cell models, plasmid replication and plasmid stability model. Dynamic simulation of batch, fed batch, steady and transient culture metabolism.

**UNIT V RECOMBINANT CELL CULTIVATION 9+3**

Different host vector system for recombinant cell cultivation strategies and advantages. E.coli, yeast *Pichiapastoris/ Saccharomyces cerevisiae*, Animal cell cultivation, plant cell cultivation, Insect cell cultivation. High cell density cultivation, process strategies, reactor considerations in the above system

**Total Hours 45+15**

*Upon Completion of this course, students will be able to get :*

**Course Outcomes**


- CO1: Analyze various operational modes of bioreactor systems*  
*CO 2: Capability to design bioreactor system for various industrial applications.*  
*CO 3: Understand and modeling different bioreactor systems with advanced modeling concepts.*  
*CO 4: Demonstrate recombinant cultivation of various plant, animal and systems for industrial applications.*  
*CO 5 understood the concept of cultivation technologies*

**Text Books**

- 1 James E. Bailey & David F. Ollis, "Biochemical Engineering Fundamentals", McGraw-Hill 2000
- 2 Anton Moser, "Bioprocess Technology", Kinetics and Reactors", Springer Verlag.1999 "
- 3 Shuler and Kargi, "Bioprocess Engineering ", Prentice Hall, 1992.

**Reference Books**

- 1 James M. Lee, "Biochemical Engineering", PHI, USA 2002.
- 2 Pauline Doran, "Bioprocess Engineering Calculation", Blackwell Scientific Publications 1998.
- 3 Harvey W. Blanch, Douglas S. Clark, "Biochemical Engineering", Marcel Decker Inc 2001.

  
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**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015  
Semester VI**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
<b>615BTT05</b>	<b>PLANT BIOTECHNOLOGY</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>	<b>50</b>	<b>50</b>	<b>100</b>

**Prerequisite** Molecular Biology

*At the end of the course, the students should be able to:*

**Course Objectives**

- *To introduce students to the principles, practices and application of plant tissue culture*
- *Learn about the transformation in science, agriculture and industry.*
- *To acquaint students with experimental design and analysis of plant biotechnology experiments.*
- *To give students hands-on experience and training in representative plant tissue culture and genetic engineering techniques.*
- *To give student wide knowledge related to practical aspects of gene transfer*

**UNIT I INTRODUCTION TO PLANT BIOTECHNOLOGY: AN OVERVIEW 9**

History of plant Biotechnology, Scope and significance of Plant Biotechnology, Plant Tissue Culture as a technique to produce novel plants and hybrids, Different types of tissue culture medium and their constituents. Plant growth hormones.

**UNIT II PLANT CELL AND TISSUE CULTURE 9**

Types of cell culture: culture of single cells, cell and organ differentiation. Stages of micropropagation. Choice of plant species for micropropagation, production of virus free plants: protoplast isolation, micropropagation work in India

**UNIT III GENE TRANSFER TO NUCLEAR GENOME 9**

Time line for utilization of gene transfer technology (event). Target cells for transformation: vector for gene transfer technology. Ti and Ri plasmids of *Agrobacterium*. Gene transfer methods: Agro-infection and gene transfer, physical delivery method.

**UNIT IV PLANT BIOTECHNOLOGY FOR AGRICULTURAL PRACTICES 9**

Biopesticides and Bioinsecticides, Integrated pest management. A total system or ecological approach of IPM. Present status and future needs for making biopesticides and IPR popular. Biofertilizers and integrated nutrients management.

**UNIT V PLANT BIOTECHNOLOGY FOR ENVIRONMENT 9**

Environment, bioenergy and biofuels, bioremediation, types of biodiversity and their applications, plant biotechnology: reasons of concern for loss of biodiversity, plant biotechnology and climate change

**Total Hours 45**

**Course  
Outcomes**

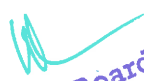
*Upon Completion of this course, students will be able to get :*  
*CO1: acquaint with principles, technical requirement, scientific and commercial applications in Plant Biotechnology,*  
*CO 2: support methodologies in plant tissue/cell culture to plant improvement, as well as DNA handling with PCR-based detection diagnostic tools,*  
*CO 3: become motivated to set goals towards pursuing graduate school and higher level positions, such as a manager and key scientist in plant biotechnological research institutes and industries.*  
*CO 4 acquired knowledge related to tissue culture*  
*CO5 gained knowledge about the gene transfer*

**Text Books**

- 1 Slater A, NW Scott, MR Fowler. Plant Biotechnology, 2nd ed. Oxford University Press, 2008.
- 2 Hopkins, W. Gand Huner, N.P.A. Introduction to Plant Physiology. 3rd ed. John Wiley & Sons Inc. New York, 2004.
- 3 Balasubramanian, Bryce, Dharmalingam, Green, Kunthalajayaraman. Concepts in Biotechnology, revised edition. Universities Press, 2007.
- 4 Karvita B Ahluwalia. Genetics. New age international Pvt. Ltd. Publishers. New Delhi. 2002.

**Reference Books**

- 1 Bhojwani and Bhatnagar. Embryology of Angiosperms, Vikar Publishing House Pvt. Ltd, New Delhi. 1981.
- 2 Sharp. Molecular Genetic Elements, Academic press, New York. 1983.

  
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**Department** BIOTECHNOLOGY      **Programme** B. TECH. - BT      **Regulation** 2015  
**Semester VI**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		CA	EA	Total
615BTP07	BIOPROCESS ENGINEERING LABORATORY -II	0	0	4	2	50	50	100

**Prerequisite** Bioprocess Engineering lab-I

*At the end of the course ,the students should be able to:*

- Course Objectives**
- To impart practical knowledge in sterilization and preparation for bioreactor
  - To develop practical knowledge of bioreactor operations in lab scale
  - To develop knowledge in mass transfer rate in bioreactor
  - To develop practical knowledge of reactor
  - To learn about the RTD process

1. Batch sterilization kinetics
2. Batch cultivation with exhaust gasanalysis
3. Estimation of KLa- Dynamic gassing out method
4. Estimation of KLa- Sulphite oxidation method
5. Estimation of KLa-Power correlation method
6. Fed batch cultivation kinetics
7. Algal cultivation
8. Residence time distribution-CSTR
9. Residence time distribution-PFR
10. Estimation of overall Heat transfer coefficient
11. Estimation of mixing time in reactor

*Upon Completion of this course, students will be able to get CO1:*

- Course Outcomes**
- Analyze various operational modes of bioreactors systems*
- CO 2: Capability to handle bioreactors system for various industrial applications.*
- CO 3: Ability to investigate, design and conduct experiments on bioprocess engineering problems*
- CO 4: ability to investigate about the RTD process*
- CO 5 understood the concept of various reactor*

**Text Books**

- 1 James E. Bailey & David F. Ollis, "Biochemical Engineering Fundamentals", McGraw-Hill 2000
- 2 Anton Moser, "Bioprocess Technology", Kinetics and Reactors", Springer Verlag.1999

**Reference Books**

- 1 James M. Lee, "Biochemical Engineering", PHI, USA 2002.
- 2 Pauline Doran, "Bioprocess Engineering Calculation", Blackwell Scientific Publications 1998.



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**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015  
Semester VI**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
615BTP08	<b>PLANT BIOTECHNOLOGY LABORATORY</b>	0	0	3	2	50	50	100

**Prerequisite** Molecular Biology

*At the end of the course ,the students should be able to:*

**Course Objectives**

- *To introduce students to the principles, practices and application of plant tissue culture and transformation in science, agriculture and industry.*
- *To acquaint students with experimental design and analysis of plant biotechnology experiments.*
- *To give students hands-on experience and training in representative plant tissue culture and genetic engineering techniques.*
- *To introduce the concept of culture techniques*
- *To give hands on experience related to embryogenesis*

1. Introduction to plant tissue culture
2. Preparation of Tissue culture medium (Murashige and Skoog)
3. Effect of Plant growth regulator of various explants for callus induction and cell suspension culture
4. *In vitro* seeds germination
5. Micropropagation of *Moringa olifera* or *Concanensis* plant by leaf disc culture
6. Organogenesis and somatic embryogenesis
7. Artificial seed preparation
8. Shoot tip and nodal sector culture
9. Callus culture
10. Meristem Culture for Virus-Free Plants
11. *Agrobacterium tumefaciens*-mediated plant transformation

*Upon Completion of this course, students will be able to get :*

**Course Outcomes**

*CO1: Understanding of the theoretical background knowledge in plant sciences needed for an understanding of plant biotechnology.*

*CO2:*

*Working knowledge of laboratory techniques used in plant biotechnology*

*CO 3: A capacity to undertake research in plant biotechnology.*

*CO 4 introduced the concept of culture techniques*


*CO 5 gain the knowledge and experience related to embryogenesis*

**Text Books**

- 1 J. Reinert and M.M. Yeoman, "Plant Cell and Tissue Culture" Springer-Verlag Berlin Heidelberg. 1982
- 2 Keith Lindsey, "Plant Tissue Culture Manual", Springer Netherlands.1997.

**Reference Books**

- 1 K. Lindsey, "Plant Tissue Culture Manual", Springer US 1992.
- 2 L.G. Nickell,, "Plant Growth Regulators", pringer-Verlag Berlin Heidelberg 1982.



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**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015**  
**Semester VI**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
615BTP09	TECHNICAL SEMINAR	0	0	2	1	50	50	100

**Prerequisite** Microbiology, Biochemistry, Molecular biology, Genetic Engineering  
*At the end of the course ,the students should be able to:*

- Course Objectives**
- To gain the knowledge of various recently developed biotechnology topics. This will help students in their PG studies


- Biochips
- Nanotechnology in medicine
- Forensic science
- Genetically modified organisms (Bt cotton and Bt brinjal etc.)
- Bioinstrumentation
- Biosensors
- Upstream process Technology
- Bioprocess Control & automation
- Biomaterials
- Protein engineering & insilico drug designs
- Artificial organs

*Upon Completion of this course, students will be able to*

**Course Outcomes** get : C01: Analyze various techniques in Biotechnology  
 Co2: Capability to handle various Instruments in the field of Biotechnology.

**Reference Books**

- James M. Lee, "Biochemical Engineering", PHI, USA 2002.
- Pauline Doran, "Bioprocess Engineering Calculation", Blackwell Scientific Publications 1998.
- Isil Aksan Kurnaz, "Techniques in Genetic Engineering" Published, CRC Press, 2015

  
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**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015  
Semester VII**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
<b>715BTT01</b>	<b>ANIMAL BIOTECHNOLOGY</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>	<b>50</b>	<b>50</b>	<b>100</b>

**Prerequisite** Genetic Engineering

*At the end of the course ,the students should be able to:*

- Course Objectives**
- *To provide the fundamentals of animal cell culture, details of the diseases and therapy*
  - *To offer the knowledge about the micromanipulation and transgenic animals*
  - *Know about transgenic animals*
  - *Learn about large scale production of animal cell cultures*
  - *Learn about the therapy activities*

**UNIT I ANIMAL CELL CULTURE 9**

Introduction to basic tissue culture techniques; Natural media, Nutritional requirement of media; chemically defined and serum free media; commonly used cell lines & their origin, various types of cultures- suspension cultures, immobilized cultures; somatic cell fusion; cell cultures as a source of valuable products; animal cell cultures and their applications, their maintenance and preservation; organ cultures. Measurement of cell viability, contact inhibition.

**UNIT II ANIMAL DISEASES, DIAGNOSIS AND THEIR THERAPY 9**

Bacterial and viral diseases in animals; diagnosis of animal diseases using monoclonal antibodies, molecular diagnostic techniques - like PCR, in-situ hybridization; northern and southern blotting, RFLP. Animal diseases; Treatment of animal diseases through recombinant cytokines, monoclonal antibodies, vaccines and gene therapy.

**UNIT III MICROMANIPULATION OF EMBRYO'S 9**

Introduction to micromanipulation technology; equipments used in micromanipulation; artificial insemination  
in vitro fertilization and embryo transfer; micromanipulation technology and intracytoplasmic sperm injection.

**UNIT IV TRANSGENIC ANIMALS 9**

Concepts of transgenic animal technology; stem cell cultures in the production of transgenic animals. DNA micro injection, lipofection, production of dolly, embryonic stem cells, retro viral method of gene insertion, calcium phosphate DNA uptake method.

**UNIT V SCALING UP OF ANIMAL CELL CULTURES 9**

Tissue culture as a screening system, cytotoxicity and diagnostic tests, mass production of important biological molecules, Harvesting of products, applications of cell culture technology in production of human and animal viral vaccines, cell culture fermenters.

*Upon Completion of this course, students will be able to get :*

**Course Outcomes**


*Co1: Understand the animal cell culture, animal diseases and its diagnosis  
Co2: Gain the knowledge of therapy of animal infections  
Co3: Know the concepts of micro manipulation technology and transgenic animal technology  
Co4 Know the concepts of micro manipulation technology and transgenic animal technology  
Co5 The concepts of transgenic animals  
Bulk production of animal cell cultures*

**Text Books**

- 1 Ranga M.M. Animal Biotechnology. Agrobios India Limited, 2002
- 2 Ramadass P, Meera Rani S. Text Book of Animal Biotechnology. Akshara Printers, 1997

**Reference Books**

- 1 Masters J.R.W. Animal Cell Culture: Practical Approach. Oxford University Press, 2000
- 2 Johnson A and Holland.A, Animal Biotechnology and ethics, Chapman & Hall Madras 1998
- 3 Culture of Animal Cells: A Manual of Basic Technique and Specialized Applications, 6th Edition, R. Ian Freshney, September 2010, Wiley-Blackwell publications

  
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**ADHIYAMAAN COLLEGE OF ENGINEERING (AUTONOMOUS), HOSUR-635 109**  
**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015**  
**Semester VII**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
715BTT02	DOWNSTREAM PROCESSING	3	0	0	3	50	50	100

**Prerequisite** Fundamentals of Unit Operation, Instrumental Methods of Analysis

*At the end of the course, the students should be able to:*

- Course Objectives**
- Understand the methods to obtain pure proteins, enzymes and in general about product development R & D
  - Have depth knowledge and hands on experience with on Downstream processes Understand the concepts in purification of biomolecules
  - Have depth knowledge in drying and crystallization
  - Gain knowledge about the finishing operation
  - Understand the method related to purification

**UNIT I DOWNSTREAM PROCESSING 9 Hrs**

Introduction to downstream processing, principles characteristics of biomolecules and bioprocesses. Cell disruption for product release- mechanical, enzymatic and chemical methods. Pretreatment of products..

**UNIT II PHYSICAL METHODS OF SEPERATION 9 Hrs**

Unit operations for solid-liquid separation: filtration- Batch and continuous filtration, Microfiltration: centrifugation- Types of centrifuge and sedimentation

**UNIT III ISOLATION OF PRODUCTS 9 Hrs**

Adsorption, liquid-liquid extraction, aqueous two-phase extraction, supercritical extraction membrane separation – ultrafiltration and reverse osmosis, dialysis, precipitation of proteins by different methods.

**UNIT IV PRODUCT PURIFICATION 9 Hrs**

Chromatography – principles, instruments and practice, adsorption, reverse phase, ion-exchange, size exclusion, hydrophobic interaction, bioaffinity and pseudo affinity chromatographic techniques, HPLC

**UNIT V FINAL PRODUCT FORMULATION AND FINISHING OPERATIONS 9 Hrs**

Crystallization – Basic Concept, Crystal size distributions, Batch Crystallization, Recrystallization. Drying – Basic concept, Drying Equipments, Conduction drying, Adiabate Drying, lyophilization of Final product **45 Hrs**

*Upon Completion of this course, students will be able to :*

- Course Outcomes**
- CO1: Ability to define the fundamentals of downstream processing for product recovery  
 CO 2: Understand the requirements of successful operations of downstream processing  
 CO 3: Describe the process of downstream equipments and explain the techniques in multifactorial manufacturing  
 CO 4: Ability to understand the knowledge in finishing operation  
 in DSP  
 CO 5: Understand the concept related to purification

### **Text Books**

- 1 P.A. Belter, E.L. Cussler And Wei-Houhu – Bioseparations – Downstream Processing For Biotechnology, Wiley Interscience Pub. (2002).  
R.O. Jenkins, (Ed.) – Product Recovery In Bioprocess Technology – Biotechnology By Open Learning Series, Butterworth-Heinemann (1998).
- 2

### **Reference Books**

- 1 E L V Harris and S. Angal, Protein Purification Methods, Ed. IRL Press at Oxford University Press, 2004.
- 2 J. E. Bailey and D. F. Ollis, Biochemical Engineering Fundamentals, 2nd Edition, Mc-Graw Hill, Inc., 2001.



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Semester VII**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		CA	EA	Total
715BTT03	CANCER BIOLOGY	3	0	0	3	50	50	100

**Prerequisite** Genetic Engineering, Molecular biology

*At the end of the course, the students should be able to:*

- Course Objectives**
- To learn about pathogenesis of cancer
  - To understand fundamentals of cancer
  - To identify cancer through tools developed by biotechnology research molecules synthesized for cancer therapy
  - To understand and preventive measure from cancer
  - To understand the activities related to metastasis

**UNIT I FUNDAMENTALS OF CANCER BIOLOGY 9 Hrs**

Introduction of cancer biology and cancer genetics, intra and extra cellular control of cell division, programmed cell death (apoptosis), intrinsic and extrinsic pathways of cell death, necrosis, malignancies, metastasis, apoptosis in relation with cancer, Regulation of cell cycle, tumour suppressor genes, different forms of cancers, diet and cancer.

**UNIT II PRINCIPLES OF CARCINOGENESIS 9 Hrs**

Theory of carcinogenesis, Chemical carcinogenesis, metabolism of carcinogenesis, Detection using biochemical assays, tumor markers, principles of physical carcinogenesis, x-ray radiation-mechanisms of radiation carcinogenesis

**UNIT III PRINCIPLES OF MOLECULAR CELL BIOLOGY OF CANCER 9 Hrs**

Signal targets and cancer, activation of kinases; Oncogenes, identification of oncogenes, retroviruses and oncogenes, detection of oncogenes. Oncogenes/proto oncogene activity.

**UNIT IV PRINCIPLES OF CANCER METASTASIS 9 Hrs**

Clinical significances of invasion, heterogeneity of metastatic phenotype, metastatic cascade, basement membrane disruption, three step theory of invasion, proteinases and Tumour cell invasion.

**UNIT V NEW MOLECULES FOR CANCER THERAPY 9 Hrs**

Different forms of therapy, chemotherapy, radiation therapy, detection of cancers, Anticancer Drugs- Classes of Anticancer Drugs, Drug Metabolism and Toxicity, Targeted Therapy in Cancer, Gene therapy.

**45**

**TotalHours**

*Upon Completion of this course, students will be able to :*

- Course Outcomes**
- CO1: The students after completing this course would be aware of cancer and causes of cancer.
- CO 2: The students would be aware of cancer detection techniques.
- CO 3: The students would be aware of identification and detection of oncogene
- CO 4: The students will come to know the new molecules used to treat cancer
- CO5: The student will come to gain knowledge related to metastasis

**Text Books**

- 1 Primrose, S.B., and R.M. Twyman, "Principles of gene manipulation and Genomics", Blackwell Publishing, MA. USA, 2006.



- 2 Twayman.R.M, "Principles of Proteomics" (Advanced text series), Taylor and Francis, 1st edition, 2004.

**Reference Books**

- 1 Brown T. A. 2007, Genomes 3. Garland Science Publishing, New York
- 2 Campbell, A.M. and Heyer, L.J., "Discovering Genomics, Proteomics and Bioinformatics", 2 nd Edition, Benjamin Cummings, 2007.
- 3 Dunham, I., "Genome Mapping and sequencing", Horizon Scientific, 2003
- 4 Read, T.D., Nelson, K.E., Fraser, C.M., "Microbial Genomes", Humana Press, Inc., USA, 2004.
- 5 Daniel C. Liebler "Introduction to Proteomics" Humana Press, 2002.

  
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Semester VII

Course Code	Course Name	Hours/Week			Credit C	Maximum Marks		
		L	T	P		CA	EA	Total
715BTT04	DISASTER MANAGEMENT	3	0	0	3	50	50	100

**Prerequisite** Environmental Science and Sustainability

*At the end of the course, the students should be able to:*

- To ensure that students begin to understand the relationship between Vulnerability, disasters, disaster prevention and risk reduction
- To gain a preliminary understanding of approaches of Disaster Risk Reduction (DRR)
- To enhance awareness of institutional processes in the country
- To develop rudimentary ability to respond to their surroundings with potential disaster response in areas where they live, with due sensitivity
- To gain concept of the disaster management

**Course Objectives**

**UNIT I INTRODUCTION TO DISASTERS**

9

Definition: Disaster, Hazard, Vulnerability, Resilience, Risks – Disasters: Types of disasters – Earthquake, Landslide, Flood, Drought, Fire etc - Classification, Causes, Impacts including social, economic, political, environmental, health, psychosocial, etc.- Differential impacts- in terms of caste, class, gender, age, location, disability - Global trends in disasters: urban disasters, pandemics, complex emergencies, Climate change- Dos and Don'ts during various types of Disasters.

**UNIT II APPROACHES TO DISASTER RISK REDUCTION (DRR)**

9

Disaster cycle - Phases, Culture of safety, prevention, mitigation and preparedness community based DRR, Structural-nonstructural measures, Roles and responsibilities of community, Panchayati Raj Institutions/Urban Local Bodies (PRIs/ULBs), States, Centre, and other stake-holders- Institutional Processes and Framework at State and Central Level- State Disaster Management Authority (SDMA) – Early Warning System – Advisories from Appropriate Agencies.

**UNIT III INTER-RELATIONSHIP BETWEEN DISASTERS AND DEVELOPMENT**

9

Factors affecting Vulnerabilities, differential impacts, impact of Development projects such as dams, embankments, changes in Land-use etc.- Climate Change Adaptation- IPCC Scenario and Scenarios in the context of India - Relevance of indigenous knowledge, appropriate technology and local resources.

**UNIT IV DISASTER RISK MANAGEMENT IN INDIA**

9

Hazard and Vulnerability profile of India, Components of Disaster Relief: Water, Food, Sanitation, Shelter, Health, Waste Management, Institutional arrangements (Mitigation, Response and Preparedness, Disaster Management Act and Policy - Other related policies, plans, programmes and legislation – Role of GIS and Information Technology Components in Preparedness, Risk Assessment, Response and Recovery Phases of Disaster – Disaster Damage Assessment.

**UNIT V DISASTER MANAGEMENT: APPLICATIONS AND CASE STUDIES AND FIELD WORKS 9**

Landslide Hazard Zonation: Case Studies, Earthquake Vulnerability Assessment of Buildings and Infrastructure:

Case Studies, Drought Assessment: Case Studies, Coastal Flooding: Storm Surge Assessment, Floods: Fluvial and Pluvial Flooding: Case Studies; Forest Fire: Case Studies, Man Made disasters: Case Studies, Space Based Inputs for Disaster Mitigation and Management and fieldworks related to disaster management.

**Total Hours 45**

*Upon Completion of this course, students will be able to :*

**Course Outcomes**

*CO 1: Differentiate the types of disasters, causes and their impact on environment and society.*

*CO 2: Assess vulnerability and various methods of risk reduction measures as well as mitigation.*

*CO 3: Draw the hazard and vulnerability profile of India, Scenarios in the Indian context, Disaster damage assessment and management.*

*CO 4: learned about the disaster management*


*CO5: variation occur with respect to atmospheric change.*

**Text Books**

- 1 Tushar Bhattacharya, "Disaster Science and Management", McGraw Hill India Education Pvt. Ltd., 2012.
- 2 Gupta Anil K, Sreeja S. Nair. Environmental Knowledge for Disaster Risk Management, NIDM, New Delhi, 2011
- 3 Kapur Anu Vulnerable India: A Geographical Study of Disasters, IAS and Sage Publishers, New Delhi, 2010.
- 4 Singhal J.P. "Disaster Management", Laxmi Publications, 2010.

**Reference Books**

- 1 Govt. of India: Disaster Management Act , Government of India, New Delhi, 2005
- 2 Government of India, National Disaster Management Policy, 2009.

  
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**ADHIYAMAAN COLLEGE OF ENGINEERING (AUTONOMOUS), HOSUR - 635 109**

**Department BIOTECHNOLOGY Programme B. TECH. - BT Regulation 2015**

**Semester VII**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
715BTP07	<b>DOWNSTREAM PROCESSING LABORATORY</b>	0	0	4	2	50	50	100

**Prerequisite** Bioprocess Engineering lab-I and Bioprocess Engineering lab -II

*At the end of the course ,the students should be able to:*

- Course Objectives**
- *To understand the nature of the end product, its concentration, stability and degree of purification required*
  - *To design processes for the recovery and subsequent purification of target biological products*
  - *To gain knowledge on sonication*
  - *To learn about the chromatography techniques*
  - *To learn about the drying equipment*

1. Solid liquid separation – Centrifugation
2. Precipitation – Ammonium sulphite precipitation
3. Aqueous two phase extraction of biologicals
4. Cell disruption techniques – Ultrasonication
5. Cell disruption techniques –Batch and continuous
6. Ultra filtration separation
7. High resolution purification – Affinity chromatography
8. High resolution purification – Size exclusion chromatography
9. High resolution purification – Ion exchange chromatography
10. Product polishing – Spray drying

*Upon Completion of this course, students will be able to get :*

*CO1: Acquired knowledge for the separation of whole cells and other insoluble ingredients from the culture broth.*

**Course Outcomes** *CO2: Learned various techniques like extraction, precipitation, membrane separation for concentrating biological products*

*CO3: Learned the basic principles and techniques of chromatography to purify the biological products and formulate the products for different end uses*

*Co4: learned about the drying equipment*


*Co5: Gained knowledge on sonication*

**Text Books**

- 1 R.O. Jenkins, (Ed.) – Product Recovery In Bioprocess Technology – Biotechnology By Open Learning Series, Butterworth-Heinemann (1992).
- 2 P.A. Belter, E.L. Cussler And Wei-Houhu – Bioseparations – Downstream Processing For Biotechnology, Wiley Interscience Pun. (1988).

**Reference Books**

- 1 J.C. Janson And L. Ryden, (Ed.) – Protein Purification – Principles, High Resolution Methods And Applications, VCH Pub. 1989.



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**Semester VII**

Course Code	Course Name	Hours/Week			Credit	Maximum Marks		
		L	T	P		C	CA	EA
715BTE10	GENOMICS AND PROTEOMICS	3	0	0	3	50	50	100

**Prerequisite** Genetic Engineering

*At the end of the course, the students should be able to:*

**Course Objectives**

- *To understand the gene cloning methods, tools and techniques involved in genome analysis and genomics.*
- *To explain the heterologous expression of cloned genes in different hosts, production of recombinant proteins and PCR techniques.*
- *To identify the importance of protein in biomolecules and the structure-function relationships in proteins.*
- *To explain comparative genomics and proteomics.*
- *To know about the functional organization of the genome and proteome*

**UNIT I ORGANIZATION OF GENOMES 8 Hrs**

Introduction: Genome, Genomics, Omics and importance, General features, C-value paradox. Gene identification; gene prediction rules and software's; Genome databases; Annotation of genome. Genome diversity: taxonomy and significance of genomes – bacteria, yeast, Caenorhabditis, Homosapiens, Arabidopsis, etc

**UNIT II MAPPING GENOMES 10 Hrs**

Genetic mapping – i) Cross breeding and pedigree analysis, ii) DNA markers - RFLPs, SSLPs, SNPs  
 Physical mapping - Restriction mapping, Fluorescent in situ hybridization, Radiation hybrid mapping and  
 Sequence tagged site mapping.

**UNIT III GENOMICS 10 Hrs**

Structural genomics: Assembly of a contiguous DNA sequence- shotgun method, clone contig method, and whole  
 –genome shotgun sequencing Understanding a genome sequence: locating the genes in a genome sequence, determining the functions of individual genes and by studying the activity of a protein coded of an unknown gene

**UNIT IV PROTEOME INFORMATICS 9 Hrs**

2D Electrophoresis - Spot visualization and picking - Database for 2D gel - Tryptic digestion of protein  
 - Peptide fingerprinting - Data analysis: Mass spectrometry; ion source (MALDI, spray sources); analyzer (ToF, quadrupole, quadrupole ion trap) and detectors - Ramachandran plot - Post-translational modification of proteins  
 - Limitation of proteomics

**UNIT V APPLICATIONS OF GENOMICS AND PROTEOMICS 8 Hrs**

Genomic medicine - Synthetic biology and bioengineering - Conservation genomics -  
 Interaction proteomics - Protein networks - Expression proteomics – Biomarkers –  
 Proteogenomics.

**Total Hours**

**45**

*Upon Completion of this course, students will be able to :*


- Course Outcomes**
- CO 1: The students after completing this course would be aware of how to clone commercially important genes and recombinant proteins.*
  - CO 2: The students would be aware of gene and genome sequencing techniques.*
  - CO 3: The students would be aware of microarrays, Analysis of Gene expression and proteomics.*
  - CO 4: To analyze the various interactions in protein makeup and different levels of protein structure.*
  - CO 5: To practice the latest application of protein science in their research.*

**Text Books**

- 1 Primrose, S.B., and R.M. Twyman, "Principles of gene manipulation and Genomics", Blackwell Publishing, MA, USA, 2006.
- 2 Twyman.R.M, "Principles of Proteomics" (Advanced text series), Taylor and Francis, 1st edition, 2004.

**Reference Books**

- 1 Brown T. A. 2007, Genomes 3. Garland Science Publishing, New York
- 2 Campbell, A.M. and Heyer, L.J., "Discovering Genomics, Proteomics and Bioinformatics", 2 nd Edition, Benjamin Cummings, 2007.
- 3 Dunham, I., "Genome Mapping and sequencing", Horizon Scientific, 2003
- 4 Read, T.D., Nelson, K.E., Fraser, C.M., "Microbial Genomes", Humana Press, Inc., USA, 2004.
- 5 Daniel C. Liebler "Introduction to Proteomics" Humana Press, 2002.



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